

# Resonant song recognition and the evolution of acoustic communication in crickets

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## Abstract

Rare behavioral phenotypes can challenge hypotheses about the evolution of the neural networks that drive behavior. In crickets, the diversity of song recognition behaviors is thought to be based on the modification of a shared neural network. We here report on a cricket with a novel resonant song recognition pattern that challenges this hypothesis. Females of the species *Anurogryllus muticus* respond to pulse patterns with the period of the male song, but also to song at twice the period. To identify the mechanisms underlying this multi-peaked recognition, we first explored minimal models of resonant behaviors. Though all of the three simple models tested (autocorrelation, rebound, resonate and fire) produced some kind of resonant behavior, only a single-neuron model with an oscillating membrane qualitatively matched the *A. muticus* behavior with regard to both period and duty cycle tuning. Surprisingly, the rebound model, a minimal model of the core mechanism for song recognition in crickets, only reproduces the behavior after inclusion of additional computations present in the song recognition network of crickets. We hypothesize that nonlinear computations, such as those leading to multi-peaked responses, can produce rapid—saltatory—behavioral change during evolution. Overall, this shows how phenotypic novelty can arise from the combination of different computations at the level of single cells and networks.

## 29 Introduction

30 Evolution has given rise to diverse animal forms and behaviors. Much of this phenotypic diver-  
31 sity is shaped by the process of mate recognition and sexual selection, and various categories  
32 of phenotypic cues—visual, acoustic, chemical, tactile—must be integrated for mate choice deci-  
33 sions to be made. For many species, acoustic signals—calling or courtship songs—are among  
34 the first features to be recognized and evaluated in mate choice decisions. The acoustic commu-  
35 nication signals produced during courtship behaviors are therefore highly diverse and contribute  
36 to species recognition. However, how the neural networks that produce this behavioral diversity  
37 evolve is largely unknown. A common hypothesis is that novel behaviors arise from shared neu-  
38 ral networks—mother networks—through small changes in connectivity and in cellular properties  
39 (Zhu et al., 2023; Bumbarger et al., 2013; Coleman et al., 2023; Ye et al., 2024; Seeholzer et  
40 al., 2018). At first sight, the idea of incremental changes in network parameters underlying be-  
41 havioral evolution is at odds with the observation that behavior can change rapidly (Gallagher  
42 et al., 2022; Xu and Shaw, 2021; Ronco et al., 2020; Yona et al., 2018) and outlier species—  
43 species with a highly unusual phenotype in a species group—challenge this mother network hy-  
44 pothesis. Evolutionary-developmental biology explains rapid morphological change—so-called  
45 "hopeful monsters"—through the re-use and modification of nonlinear gene-regulatory modules  
46 (Goldschmidt, 1940; Müller, 2007). Because of these nonlinearities, large morphological change  
47 can then arise from a single mutation in a saltatory instead of gradual manner. Similarly, behav-  
48 ioral innovations—"behavioral monsters"—could emerge from small changes in a neural network  
49 from the nonlinear mapping between the network's parameters and the behavior.

50 Experimental tests of the mother network hypothesis are challenging, because they involve  
51 characterizing and comparing the network properties across many species in a group and then  
52 causally linking the changes in network properties to changes in behavior. However, a precon-  
53 dition for the mother network hypothesis is that the shared network has the capacity to produce  
54 the diverse species-specific behaviors in a group. Computational modeling can help assess this  
55 capacity from behavioral data, by comparing the observed behavioral diversity with that produced  
56 by a computational model of the shared mother network. If the model of the proposed mother net-  
57 work fails to reproduce the behavior of a specific species then that species likely has undergone  
58 more drastic changes in its recognition mechanism inconsistent with the mother network hypothe-  
59 sis. Conversely, the hypothesis is supported if the network can reproduce all observed behaviors,  
60 including those of the "behavioral monsters"—species with unusual behavioral phenotypes.

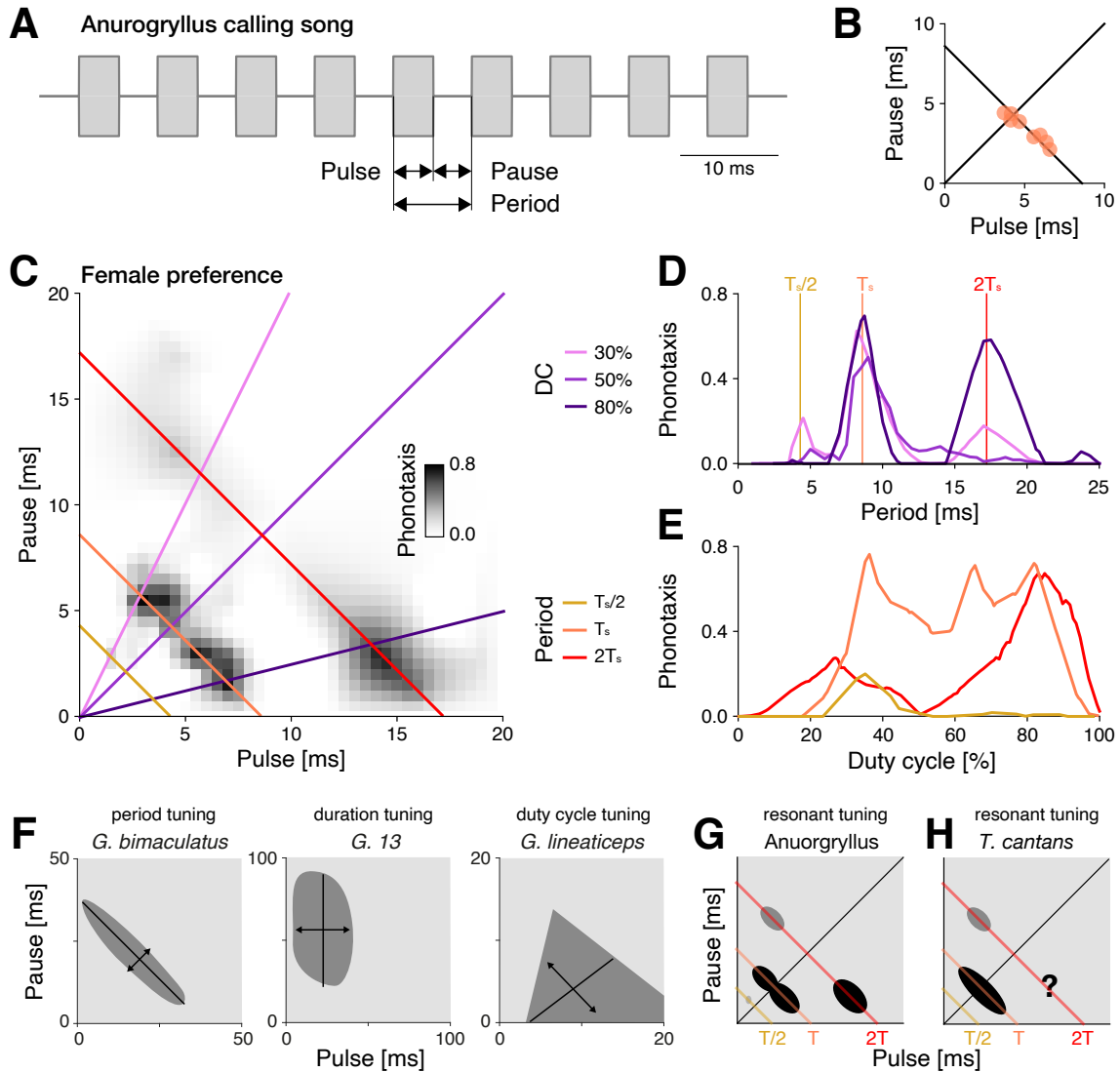
61 We address the question of behavioral diversity and neuronal evolution in the context of acous-  
62 tic communication in crickets. Males produce pulsed calling songs with species-specific pulse and  
63 pause durations ranging between 10 and 80 ms (Fig. 1A, B) (Alexander, 1962). The songs are  
64 either produced in chirps consisting of a few pulses or continuously, in trills. Females evaluate  
65 the song on the time scale of pulse pause and duration, and of chirps/trills (Fig. 1A) (Grobe et al.,  
66 2012). Attractive songs elicit positive phonotaxis in the female. The female tuning for the calling  
67 songs can be quantified by measuring the phonotactic behavior for artificial pulse patterns in a  
68 two-dimensional parameter space spanned by pulse and pause duration (Fig. 1C). The strength  
69 of phonotactic orientation towards the acoustic stimulus then serves as a measure for the strength  
70 of recognition. So far, preference functions are known from 18 cricket species, and they all reveal  
71 unimodal preferences for a single continuous range of song features (Bailey et al. (2017), Cros and  
72 Hedwig (2014), Gray et al. (2016), Hennig (2003), Hennig (2009), Rothbart and Hennig (2012a),  
73 Rothbart and Hennig (2012b), Hennig et al. (2016), and Blankers et al. (2015) and Ralf Matthias  
74 Hennig, unpublished data). The known preferences fall into three types, characterized by the fe-  
75 males' selectivity for specific features of the pulse song: Tuning for pulse duration, for period (pulse  
76 plus pause) and for duty cycle (duration divided by period, referred to from now as "DC") (Fig. 1F).  
77 Tuning for pause duration is a fourth possible phenotype, but this one has not yet been reported in  
78 crickets. Song recognition based on the duration or period of acoustic signals is not restricted to  
79 crickets but found throughout the animal kingdom (Baker et al., 2019; Araki et al., 2016; Perrodin  
80 et al., 2023; Lameira et al., 2024). Understanding the principles underlying the evolution of pulse  
81 song recognition in crickets can therefore inform similar studies in others species groups.

82 We have recently shown that the song recognition network described in the period-tuned *Gryl-*  
83 *lus bimaculatus* can produce the diversity of song recognition known in crickets until now. In *G.*  
84 *bimaculatus*, five neurons recognize the song in five steps Schöneich et al., 2015: 1) The ascend-  
85 ing neuron 1 (AN1) pools and transmits to the brain information from auditory receptors in the  
86 prothorax and produces an intensity-invariant copy of the song pattern (Benda and Hennig, 2008).  
87 2) The local neuron 2 (LN2) receives input from AN1 and provides inhibition to LN5 and LN4. 3)  
88 The non-spiking LN5 produces a post-inhibitory rebound potential at the end of each song pulse.

89 4) LN3 fires only in response to coincident input from the rebound in LN5 and a delayed input from  
90 AN1. The input delay from AN1 is tuned such that coincidence only occurs for pulses with the  
91 species-specific period of 30 ms. 5) LN4 receives inhibition from LN2, which further sharpens the  
92 feature tuning. The tuning of LN4 for pulse song matches that of the phonotaxis response. Similar  
93 principles of temporal pattern recognition with delay lines and post-inhibitory rebounds are known  
94 from many systems (Carr and Konishi, 1988; Carr, 1993; Kopp-Scheinflug et al., 2011) and un-  
95 derstanding the capacity and constraints of this algorithm in crickets can therefore shed light on  
96 temporal pattern recognition across systems. A computational model reproduced the response  
97 dynamics of all neurons in this network as well as the behavioral output (Clemens et al., 2020)  
98 revealed that the network from *G. bimaculatus* can produce the three preference types known in  
99 crickets—preference for period, pulse duration, and duty-cycle—through changes in network pa-  
100 rameters like synaptic strengths or intrinsic neuronal properties. Thus, the *G. bimaculatus* network  
101 could be the mother network producing the diversity of song recognition in crickets.

102 We here describe the male song and female preference of a novel cricket species *Anurogryl-*  
103 *lus muticus* (from now referred to as *Anurogryllus*). *Anurogryllus* females exhibit a multi-peaked  
104 recognition phenotype that is unique in crickets and could challenge the hypothesis of a shared  
105 mother network: Females are attracted not only to the period of the male song but also to twice the  
106 period (Fig. 1C–E). Importantly, all other known cricket species have preference functions with a  
107 single peak (Bailey et al. (2017), Cros and Hedwig (2014), Gray et al. (2016), Hennig (2003), Hen-  
108 nig (2009), Rothbart and Hennig (2012a), Rothbart and Hennig (2012b), Hennig et al. (2016), and  
109 Blankers et al. (2015) and Ralf Matthias Hennig, unpublished data) and only *Anurogryllus* exhibits  
110 this multi-peaked preference function. All existing evidence therefore points towards *Anurogryllus*  
111 having a phenotype that is highly unusual and an outlier in the context of crickets, consistent with  
112 the concept of “behavioral monsters”. Responses to multiples or fractions of a song’s period have  
113 only been shown in a single species of katydids, *Tettigonia cantans* (Fig. 1H), and such responses  
114 are consistent with a resonant mechanism for song recognition (Bush and Schul, 2006). Compu-  
115 tational modeling in katydids has suggested that delay-based mechanisms can not explain the  
116 resonant responses in the katydid and provided evidence for a nonlinear resonant-and-fire (R&F)  
117 mechanism of song recognition in katydids (Webb et al., 2007). Importantly, it is unclear whether  
118 the computational model of the song recognition network in crickets—which relies on a delay-  
119 based mechanism—can produce the resonant preference of *Anurogryllus*. Thus, *Anurogryllus* is  
120 a challenge to the mother network hypothesis and an opportunity to identify the computational  
121 principles that can give rise to resonant tuning.

122 Here, we provide further support for the mother network hypothesis, by demonstrating that it can  
123 produce the resonant recognition behavior of *Anurogryllus*. We first explore the tuning properties  
124 of minimal models of resonant behavior based on network and intracellular mechanisms, and  
125 compare these results to those obtained from the full mother network model. Lastly, we explore  
126 the hypothesis that nonlinear computations, such as those that give rise to resonant recognition  
127 behaviors, could form the substrate for saltatory behavioral evolution.



**Figure 1: Anurogryllus is a cricket species with resonant song recognition.**

**A** Schematic of the calling song of males from the cricket species *Anurogryllus muticus* (from now referred to as *Anurogryllus*). The song consists of a train of pulses with a specific pulse duration and pause. The period is the sum of pulse and pause and corresponds to the song's rhythm. The duty cycle (DC) is the percentage of the period occupied by the pulse and corresponds to the song's energy.

**B** Pulse and pause parameters from eight *Anurogryllus* males. The diagonal line corresponds to a DC of 50%, the anti-diagonal to the average pulse period  $T_s = 8.6$  ms. See Table 1 for all song parameters.

**C** Female phonotaxis for pulse trains with different duration and pause parameters visualized as a pulse-pause field (PPF). Phonotaxis is color coded with darker greys, representing stronger phonotactic responses (see color bar). Diagonal lines indicate stimuli with DCs of 30, 50, and 80%, shown in D as the phonotaxis along these diagonals. The anti-diagonal lines show transects with constant period stimuli shown in E at the average pulse period of the male song  $T_s$  (orange), at half ( $T_s/2$ , yellow) and twice ( $2T_s$ , red) the song period. Females respond strongly to pulse patterns with the period of the males' song, but also at twice that period, indicating resonant song recognition. See table S6 for statistical significance of the individual peaks. The PPF was obtained by interpolation of the average phonotaxis values measured for 75 artificial stimuli in 3–8 females (Fig. S1).

**D** Period tuning as a function of DC given by three transects through the PPF in C (see legend in C). Vertical lines indicate  $T_s/2$  (yellow),  $T_s$  (orange), and  $2T_s$  (red).

**E** DC tuning as a function of song period, derived from transects through the PPF in C (see legend in C).

**F** The three previously known female preference types for the pulse pattern of the male calling song in different cricket species: period (left), duration (middle), and DC (right). The solid black lines indicate the major or most tolerant axis that defines the tuning type, and the double sided arrows perpendicular to the major axis show the most sensitive feature axis.

**G** Schematic of the novel resonant recognition from *Anurogryllus*, simplified from C.

**H** Resonant recognition from the katydid *Tettigonia cantans* (Bush and Schul, 2006). The question mark indicates the range of stimulus parameters not tested in the original study. Anti-diagonal lines in G and H indicate stimuli with  $T_s/2$  (yellow),  $T_s$  (orange), and  $2T_s$  (red).

See also Fig. S1 and Table S6.

## Results

### Anurogryllus exhibits an unusual resonant recognition phenotype

The calling song of *Anurogryllus* males consists of continuous trills with a pulse period  $T_s$  of  $8.5 \pm 0.3$  ms, which corresponds to a pulse rate  $f_s$  of  $117.1 \pm 4.3$  pulses per second (Fig. 1A, B). We refer to the pulse rate measured in pulses per second as  $f$ , for notational simplicity. This pulse rate is unusually high for cricket songs, which have pulse rates between 10 and 50 pulses per second (Weissman and Gray, 2019). The song's DC—given by the ratio of pulse duration and pulse period, and indicating how much of the song is filled by pulses—is  $60 \pm 10\%$  (see Table 1 for a list of all song parameters). To quantify the preference of *Anurogryllus* females for the calling song we quantified the strength of the females' phonotaxis response during playback of 75 artificial pulse trains with different pulse and pause duration combinations (Fig. 1C, S1). This confirms that females are attracted (perform positive phonotaxis) to the pulse trains produced by conspecific males: The two-dimensional preference function spanned by pulse duration and pause contains a broad peak covering periods of 8.5 ms and DCs of 33–80%, which overlaps with the distribution of male songs. This peak is partially split along the DC axis (Fig. 1C).

Song parameter	Mean $\pm$ std	Range
Carrier frequency	7.0 $\pm$ 0.3 kHz	6.5-7.5 kHz
Pulse duration	5.1 $\pm$ 1.0 ms	3.7-6.6 ms
Pulse pause	3.4 $\pm$ 0.8 ms	2.1-4.4 ms
Pulse period	8.5 $\pm$ 0.3 ms	8.1-9.0 ms
Pulse rate	117 $\pm$ 4 pulses/s	111-124 pulses/s
Pulse DC	60 $\pm$ 10 %	50-80 %

**Table 1: Parameters of the calling song of *Anurogryllus* males.**

Data from 8 males over 1 minute of song with at least 7500 pulses per male. Carrier frequencies from (Erregger et al., 2017).

However, the phonotaxis experiments also reveal that females are attracted to songs that differ substantially from the conspecific song and the tuning of these off-target responses implies a resonant recognition phenotype in *Anurogryllus* (Fig. 1D, E, Table 1). These off-target responses appear at twice or half the song period: First, song with twice the period of the male song (17 ms) with a high DC (90%) is almost as attractive as the conspecific song. Second, females are also weakly attracted to song with twice the conspecific period (17 ms) and lower DC (25%). Lastly, there is a weak and non-significant response peak at half the conspecific period (4.5 ms) and low DC (33%). The responses at integer fractions or multiples of the song's fundamental rate indicate a resonant response mechanism. If we define  $T_s = 8.6$  ms as the period of the male song, and the fundamental rate  $f_s = 1/T_s = 116$  pulses per second, then the weak peak at half the period,  $T_s/2 \approx 4.3$  ms, corresponds to the second harmonic,  $2f_s$ , while the peaks at twice the period,  $2T_s \approx 17.2$  ms, correspond to the second subharmonic,  $f_s/2$ .

So far, a resonant song recognition behavior—with responses to three different types of pulse patterns—has not been reported in a cricket (Fig. 1F–G)—it was previously known only in the katydid *Tettigonia cantans* (Bush and Schul, 2006) (Fig. 1H). The resonant phenotype in *T. cantans* is similar to that of *Anurogryllus*: *T. cantans* females are attracted to pulse trains with the period of the male song (period 40 ms, DC 50%), and to subharmonics of the male song—songs with twice the conspecific period (80 ms, DC 25%). *T. cantans* does not respond to harmonics (half the period, 20ms) and it was not tested whether females are attracted at twice the period at higher DCs, the pattern that *Anurogryllus* is most responsive to apart from the conspecific song. A simple delay-line based mechanism in *T. cantans* was ruled out as a potential mechanism for resonance using experimental tests, but a resonate-and-fire neuron model with oscillatory membrane properties could reproduce the resonant song preference (Bush and Schul, 2006; Webb et al., 2007). Oscillatory neurons have therefore been proposed as a mechanism for song recognition in *T. cantans*. However, the rebound-based mechanism at the core of the song recognition network in crickets had not been considered, and it is unclear whether oscillatory neurons can reproduce the particular pattern of resonance observed in *Anurogryllus*.

## Simple models provide insight into the computational mechanisms of resonant tuning

The resonant phenotype in *Anurogryllus* challenges the mother network hypothesis, as the model of the song recognition network in crickets was only shown to produce all known single-peaked phenotypes, not the specific resonant phenotype of *Anurogryllus* (Clemens et al., 2021) (Figs 1F, G). We therefore tested whether this model network could also produce the resonant tuning of *Anurogryllus*. However, given that the computational model of the song recognition network in crickets is complex and has many parameters, we decided to first identify the computational principles and constraints that shape resonant tuning by investigating the ability of simple network and single-neuron models to qualitatively reproduce the resonant behavior of *Anurogryllus*. Simple models allow us to 1) isolate the minimal set of computations required for generating resonant behaviors, 2) facilitate the interpretation of the more complex network model, and 3) rule in or out alternative mechanisms not currently part of the mother network but that might be easily acquired during evolution. Given the simplicity of the models chosen, our goal was not a detailed reproduction of the *Anurogryllus* behavior (Fig. 1C), but a reproduction of the most prominent properties of the period and DC tuning: namely the broad DC peak at the period of the male song,  $T_s$ , and the two response peaks at  $2T_s$ , with the dominant peak at high DC (Fig. 1G).

We fitted three simple models to the behavioral data from *Anurogryllus*: First, an autocorrelation model, which consists of a delay line and a coincidence detector (Bush and Schul, 2006). This is the simplest model that can produce resonances and shows that delays alone can produce resonant response peaks. Second, the rebound model, which is an extension of the autocorrelation model and captures the core computation of the mother network, in which the non-delayed input to the coincidence detector consists of offset responses from a post-inhibitory rebound (Schöneich et al., 2015; Clemens et al., 2021). The rebound model will reveal whether the core-computation of the mother network—a delay line, rebound, and coincidence detection—is sufficient to produce the resonant tuning of *Anurogryllus*. Lastly, we examined the a resonate-and-fire (R&F) neuron, a single-neuron model with subthreshold membrane oscillations that reproduced the resonant behavior of *T. cantans* (Izhikevich, 2001; Webb et al., 2007). This last model will allow us to examine how changes in intracellular properties, rather than network properties, can produce the resonant song recognition of *Anurogryllus*.

### Autocorrelation models produce resonant tuning but do not match the *Anurogryllus* behavior

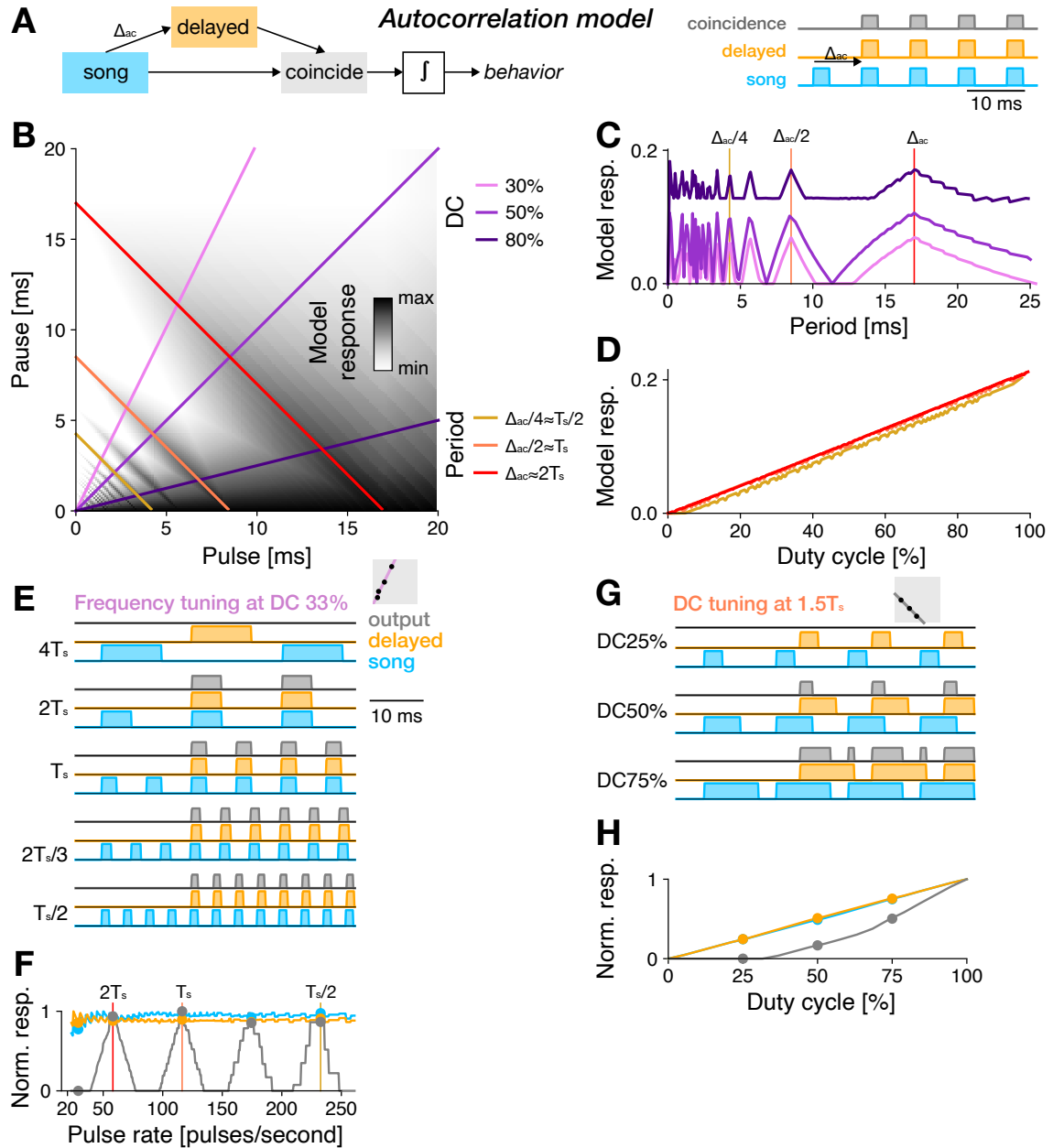
In an autocorrelation model, the song input is split into two pathways, one with a delay  $\Delta_{ac}$ , and one without a delay (Fig. 2A). Responses from the delayed and non-delayed pathways are then multiplied in a coincidence detector, that only responds when the delayed and the non-delayed inputs overlap in time. The model response is then taken as being proportional to the average output of the coincidence detector over the song.

The autocorrelation model fitted to the *Anurogryllus* data produces resonant response peaks for pulse rates at integer fractions, but not at multiples, of the delay  $\Delta_{ac}$  (Fig. 2B, C). The fitted value of  $\Delta_{ac} = 17$  ms corresponds to  $2T_s$ , the peak at twice the pulse period in the behavioral data (Fig. 1E). Coincidence occurs if  $nT = \Delta_{ac}$ , leading to resonant peaks at periods that are fractions of the delay  $T = \Delta_{ac}/n$  (or at pulse rates  $f = n/\Delta_{ac}$ ) (Fig. 2E, F). Thus, resonant peaks in the autocorrelation model arise at even and odd fractions of  $\Delta_{ac}$  and coincide with  $T_s$  and  $2T_s$ . However, the behavior only exhibits responses at even fractions of  $\Delta_{ac}$ . The lack of peaks at odd fractions of  $\Delta_{ac}$  in *Anurogryllus* renders a pure autocorrelation-based mechanism for song recognition unlikely (Fig. 1E).

Similar to the period tuning, the DC tuning of the fitted autocorrelation also does not match the behavioral data: The output of the autocorrelation model increases linearly with DC (Fig. 2D), with maximal responses for constant tones without a pause (DC 100%). By contrast, *Anurogryllus* exhibits complex DC tuning with multiple peaks and, importantly, does not respond well to pulse trains with very high DCs (Fig. 1E). The DC bias in the autocorrelation model arises because songs with longer pulses and shorter pauses are more likely to produce coincidence for any given delay (Fig. 2F, Fig. 2G, H).

In sum, the autocorrelation model demonstrates that a delay is sufficient to produce resonance. However, autocorrelation alone is insufficient to qualitatively reproduce the pulse rate and DC tuning found in *Anurogryllus*.





**Figure 2: An autocorrelation model produces resonant tuning.**

**A** In the autocorrelation (AC) model, a non-delayed (blue) and delayed (orange) copy of the stimulus are multiplied in a coincidence detector (grey). The output of the coincidence detector is integrated over the stimulus to predict the model response. The example traces show coincidence for song with a pulse period that equals the delay  $\Delta_{ac}$ .

**B** PPF for the autocorrelation model fitted to the preference data in 1C. Predicted response values are coded in greyscale (see color bar). Colored lines correspond to the DC and period transects shown in C and D (see legend).

**C** Period tuning of the autocorrelation model for different DCs (see legend in B). Resonant peaks arise at even and odd fractions of the delay parameter  $\Delta_{ac} \approx 2T_s$ . Vertical lines indicate the pulse periods transects shown in B.

**D** DC tuning for three different pulse periods (see legend in B), corresponding to  $T_s/2$ ,  $T_s$ , and  $2T$ . DC tuning is high-pass for all periods.

**E** Response traces from the autocorrelation model for songs with different periods (fractions and multiples of  $T_s$ ) and a DC of 33%. Resonant peaks arise from coincidence at integer fractions (e.g.  $1\Delta_{ac}/2 = 2T_s/3$ ) but not at multiples ( $2\Delta_{ac} = 2T_s$ ) of the delay parameter (stimulus—blue, delayed stimulus—orange, response—grey, see legend to the right).

**F** Pulse rate tuning given by the integral of the stimulus (blue), the delayed stimulus (orange), and the response (grey) at 50% DC. Response peaks arise at integer multiples and fractions of  $\Delta_{ac}$ . Dots indicate pulse patterns shown in E. Vertical lines indicate the song periods shown in D.

**G, H** Response traces for different DCs (25, 50, 75%) (G) and DC tuning (H) at a non-resonant pulse rate ( $1.5T_s = 12.9$  ms). Increasing the DC leads to coincidence even at this non-resonant pulse rate. Same color code as in E, F.

Gray boxes in E and G illustrate the stimulus parameters for which traces are shown in the context of the PPF (compare B).

## 226 **A rebound mechanism suppresses responses to pulse trains with high duty cycles**

227 The core computation for song recognition in the cricket *G. bimaculatus* is an extension of the  
228 autocorrelation model (Schöneich et al., 2015; Clemens et al., 2021) (Fig. 3A): As in the au-  
229 tocorrelation model, the song is split into a delayed and a non-delayed path. The non-delayed  
230 path is then sign-inverted and filtered to produce transient responses at the end of each pulse,  
231 to mimic a post-inhibitory rebound. The rebound model produces outputs only when the delayed  
232 input coincides with the rebound.

233 The pulse rate tuning of the rebound model resembles that of the autocorrelation model, with  
234 resonant peaks arising close to even and odd fractions of the delay  $\Delta_{rb}$  (Fig. 3B, C, compare  
235 2C). However, the fitted value of  $\Delta_{rb} = 23\text{ms}$  matches neither multiples nor fractions of  $T_s$ . This  
236 is because the rebound is produced at the end of each pulse and coincidence therefore occurs  
237 if  $n \cdot T + D = \Delta_{rb}$ , where  $D$  is the pulse duration (Fig. 3E, F). Resonant peaks occur at  $T =$   
238  $(\Delta_{rb} - D)/n$  (equation ) or  $f = n/(\Delta_{rb} - D)$ , close to even and odd fractions of  $2T_s$  (Fig. 3A, B,  
239 E, F). The responses to odd fractions of  $T_s$  in the rebound model are not found in the behavioral  
240 data. Therefore, a pure rebound mechanism is unlikely to produce the Anurogryllus behavior.

241 The DC tuning of the rebound model is band-pass, with reduced but non-zero responses for  
242 continuous tones (high DC) (Fig. 3D, G). This band-pass tuning arises from two opposing pro-  
243 cesses: On the one hand, responses increase with pulse duration up to a point set by the duration  
244 of the inhibitory filter lobe that produces the rebound. This is because the rebound is strongest  
245 if the pulse is long enough to saturate the rebound, which happens when it fully overlaps the in-  
246 hibitory filter lobe (Fig. 3I). However, a further increase in pulse duration at a fixed pulse period  
247 shortens the pauses and for short pauses, the rebound is interrupted by the next pulse (Fig. 3G,  
248 J).

249 Overall, the rebound model fails to reproduce the qualitative features of the Anurogryllus re-  
250 sponses. Period tuning exhibits excess peaks at odd fractions of the pulse rate as in the autocor-  
251 relation model. While the DC tuning is band-pass, as in Anurogryllus, responses to constant tones  
252 are still evident and the characteristic pattern with split-peaks is missing. This failure to reproduce  
253 the Anurogryllus behavior is surprising given that the rebound constitutes the core mechanism of  
254 song recognition in crickets. However, we will show below that a rebound mechanism can produce  
255 the Anurogryllus behavior when combined with other computations found in the full network.

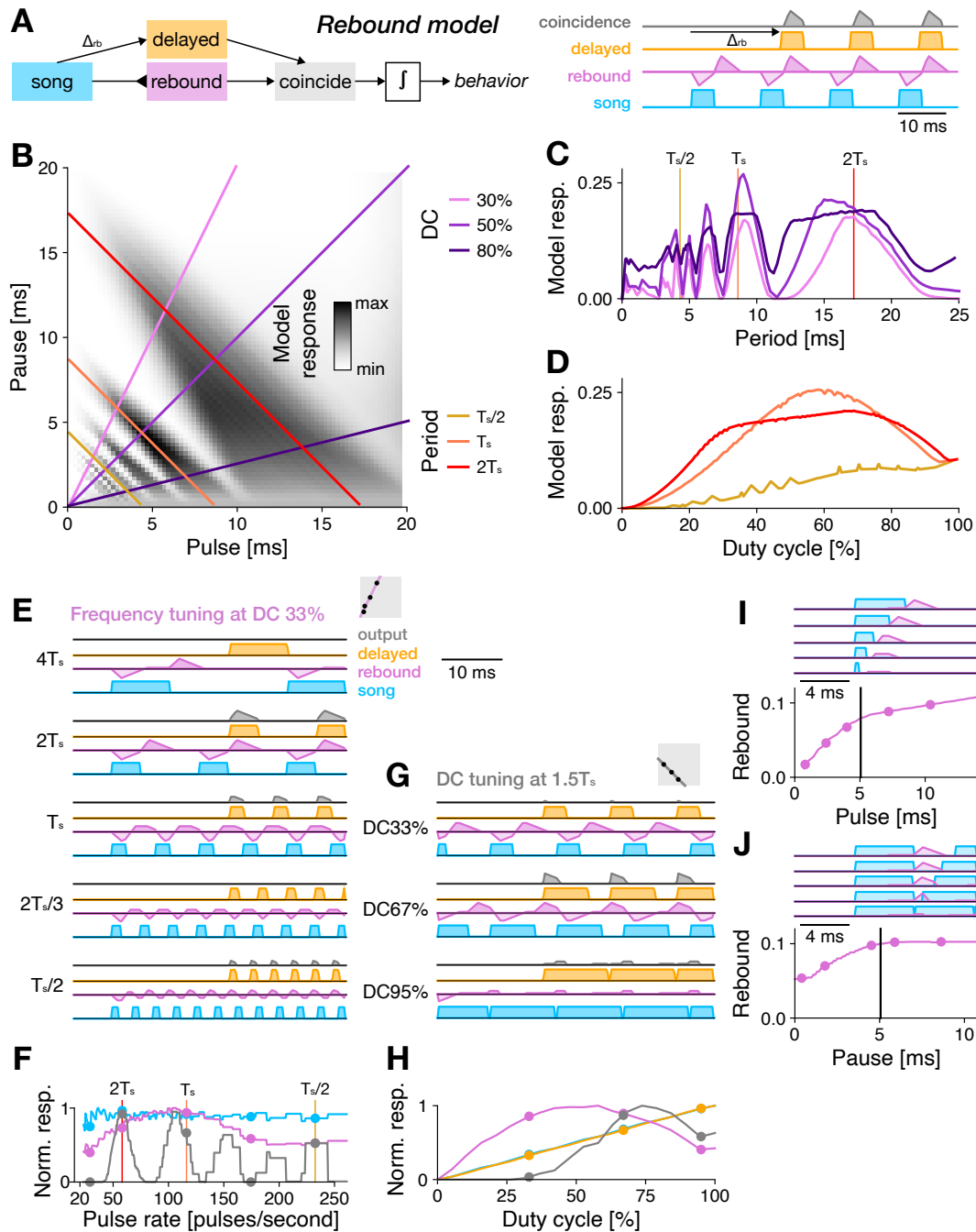
## 256 **The resonate and fire model is a simple model that qualitatively matches the Anurogryllus** 257 **behavior best**

258 As the last simple model, we fitted a resonate-and-fire (R&F) model to the Anurogryllus data. In  
259 contrast to the autocorrelation and rebound models, which are network models, the R&F model  
260 is a single neuron model that consists of coupled current and voltage-like variables (Fig. 4A)  
261 (Izhikevich, 2001). This coupling leads to input-driven damped oscillations with a characteristic  
262 frequency  $f_{r\&f}$ . Inputs that arrive at positive/negative phases of the oscillation amplify/suppress  
263 this oscillation. If the voltage reaches a threshold, a spike is elicited and the current and voltage  
264 are reset. The R&F model can produce resonant behavior if the damping is weak and it was used  
265 to reproduce the resonant song recognition from *T. cantans* (Bush and Schul, 2006; Webb et al.,  
266 2007).

267 The R&F model fitted to Anurogryllus data is weakly damped (less than 2% of the stimulus  
268 gain). It has a characteristic frequency  $f_{r\&f} = 109$  Hz, which translates to  $T_{r\&f} = 9.2$  ms—close  
269 to the pulse period of the Anurogryllus song. The R&F responds strongly if the incoming pulses hit  
270 the intrinsic oscillation during excitatory phases and resonant peaks therefore arise at multiples of  
271 the period  $n \cdot T_{r\&f} = n/f_{r\&f}$ . Thus, contrary to the autocorrelation and rebound models, the R&F  
272 model responds only to multiples (subharmonics), but not to fractions of  $T_{r\&f}$  (harmonics) (Fig. 4B,  
273 C). In the R&F, responses to fractions of  $T_{r\&f}$  are suppressed, because inputs faster than  $T_{r\&f}$  will  
274 arrive not only during the excitatory but also during the inhibitory phase of the intrinsic oscillation,  
275 reducing the net drive to the spike generator. By contrast, responses at multiples of  $T_s$  exist  
276 because subsequent pulses will arrive during the excitatory phase of the membrane oscillation  
277 (Fig. 4E, F). The R&F model produces the Anurogryllus responses at  $T_s$  and  $2T_s$  and, apart from  
278 excess responses at higher multiples of  $T_s$ , matches the behavior well.

279 The DC tuning of the R&F model is more complex than that of the autocorrelation and rebound  
280 models. At  $T_{r\&f}$ , the model responds with a single, broad peak to different DCs, whereas at  $2T_{r\&f}$ ,  
281 two separate peaks—at high and low DCs—are visible (Fig. 4B, D). The peak at the higher DC is  
282 greater than that at the lower DC, consistent with the Anurogryllus behavior. With this DC tuning,  
283 the R&F model qualitatively matches the Anurogryllus data best out of all models tested so far (Fig.





**Figure 3: Tuning for pulse rate and duty cycle in the rebound model fitted to Anurorgryllus behavior.**

**A** The rebound model is an extension of the autocorrelation model. The non-delayed branch (purple) is sign-inverted (blunt ended arrow indicates inhibition) and filtered by a bi-phasic filter to produce transient responses at pulse offsets that mimic a post-inhibitory rebound. The positive part of the rebound and the delayed stimulus are then combined through coincidence detection.

**B** PPF for the rebound model fitted to the preference data in 1C. Predicted response values are color coded (see color bar). Colored lines correspond to the DC and period transects shown in C and D (see legend).

**C** Period tuning of the rebound model for different DCs (see legend in C). Vertical lines correspond to the pulse period transects shown in B.

**D** DC tuning for three different pulse periods (see legend in C). DC tuning is high-pass for short periods ( $T_s/2$ , yellow) and band-pass for intermediate and long periods ( $T_s$  (orange),  $2T_s$  (red)).

**E** Response traces of the rebound model for songs with different periods (fractions and multiples of  $T_s$ ) and a DC of 33% (stimulus—blue, rebound response—pink, delayed stimulus—orange, response—grey, see legend to the right).

**F** Pulse rate tuning given by the integral of the stimulus. Response peaks arise at integer multiples of  $f$ . Dots indicate periods shown in E.

**G, H** Response traces for a DC sweep (33, 67, 95%) (G) and DC tuning (H) at a non-resonant period of  $1.5T_s = 12.9$  ms. Even at this non-resonant period, responses increase with DC, consistent with the broadening of the response peaks with DC in B and C. Responses decrease at very high DCs (short pauses), because the rebound is truncated by the next pulse (see J). Same color scheme as in E, F.

**I, J** Integral of the rebound as a function of pulse duration (I) and pause (J). Dots in the curves (bottom) indicate example traces shown on top of each curve. A minimum pulse duration and pause duration (black lines) are required for the rebound to fully develop. At short pauses the rebound is interrupted by the following pulse (J).

Gray boxes in E and G illustrate the stimulus parameters for which traces are shown in the context of the PPF (compare B).

284 1F). Note that the R&F also produces peaks at  $3T_{r\&f}$ , but stimuli covering these periods were not  
285 tested experimentally.

286 How does this complex DC tuning arise in the relatively simple R&F model? In the model,  
287 inputs during the excitatory phase of the membrane potential oscillation amplify the oscillation and  
288 therefore elicit spiking responses, while inputs during the negative phase suppress the spiking  
289 responses. Songs with a pulse period of  $T_s$  match the period of the membrane oscillation and  
290 an input with a DC of 50% will produce the maximum output because it covers only the excita-  
291 tory phase of the oscillations (Fig. 1G, H). Shorter pulses (DC<50%) will produce weaker voltage  
292 responses because they engage the excitatory phase less, and longer pulses (DC>50%) will pro-  
293 duce weaker voltage responses because they extend into the inhibitory phase. Pulse patterns  
294 with a pulse period of  $2T_s$ —twice the period of the oscillation—produce DC tuning with two broad  
295 peaks—around DC 25% and around DC 75%—and no responses at DC 50% (Fig. 1I, J). The  
296 responses at DC 50% are suppressed because the pulse covers one full period of the oscillation,  
297 and therefore equally engages the excitatory and the inhibitory phases of the oscillation, resulting  
298 in weak spiking responses. Stimuli with smaller or larger DCs produce stronger responses be-  
299 cause more of the excitatory phases of the oscillation are engaged. The peak at higher DCs is  
300 higher than that at lower DCs because the pulse hits the excitatory phase once per period for DCs  
301 below 50% and twice for DCs above 50% (Webb et al., 2007), as in (Fig. 4I).

### 302 **Simple network models, unlike the single neuron model, fail to reproduce the behavioral** 303 **period and DC characteristics**

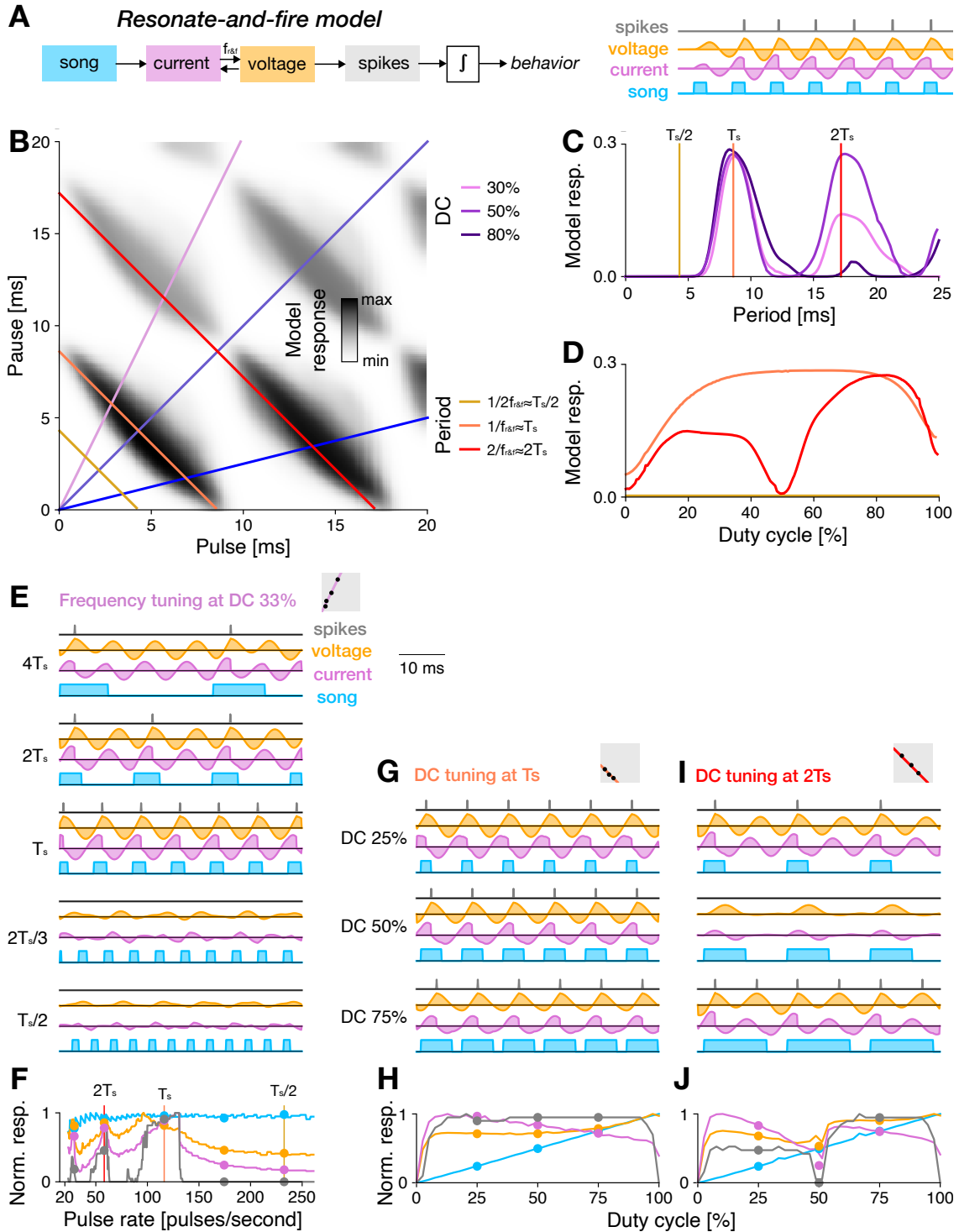
304 Overall, none of the simple models were able to fully reproduce the Anurogryllus tuning. However,  
305 a single-neuron model—the R&F model—came closest, suggesting that changes in single neuron  
306 properties might underlie the emergence of resonant tuning in Anurogryllus (Fig. 4). By contrast,  
307 simple delay-based models (autocorrelation and rebound) are insufficient to recover the Anurogryl-  
308 lus tuning (Figs 2, 3): The delay-based models are resonant but they produce strong responses  
309 to very short periods (fractions of  $T_s$ ) and are unable to replicate the DC tuning of Anurogryllus,  
310 in particular the double-peaked DC tuning at  $2T_s$ . Importantly, the failure of the rebound model,  
311 which replicates the hypothesized core mechanism of song recognition in crickets, challenges the  
312 mother network hypothesis (Schöneich et al., 2015; Clemens et al., 2021)). However, the mother  
313 network, developed using electrophysiological data from *G. bimaculatus*, contains additional com-  
314 putations like adaptation and feed-forward inhibition. We therefore fitted a model of the full network,  
315 previously developed in Clemens et al. (2021) (Fig. 5A), to the behavioral data from Anurogryllus  
316 to test whether these additional computations can produce the behavior.

### 317 **The mother network can produce the resonant phenotype**

318 A computational model of the song recognition network in crickets, that was originally constructed  
319 to reproduce electrophysiological data from *G. bimaculatus* (Clemens et al., 2021), was fitted to the  
320 behavioral data from Anurogryllus females (Fig. 1C). This model reproduced the Anurogryllus be-  
321 havior (5A): Resonant peaks at  $T_s$  and  $2T_s$ , and DC tuning at  $2T_s$  that is bimodal with a preference  
322 for higher DCs. This supports the mother network hypothesis—the network from *G. bimaculatus*  
323 can produce the preference profiles from all cricket species examined so far and could therefore  
324 constitute the template network for song recognition in crickets. How does the characteristic period  
325 and DC tuning arise in the network? Above, we have shown that the rebound mechanism at the  
326 core of the network is sufficient to produce resonant responses but insufficient to produce the DC  
327 tuning at  $2T_s$  (Fig. 3 D). We therefore investigated where in the network both response properties  
328 arise.

329 In the full network, LN3 is equivalent to the output of the simple rebound model as it is the  
330 coincidence detector that receives input from the rebound neuron LN5 and a delayed input from  
331 AN1. Accordingly, resonant tuning with responses at multiple periods in the network arises in LN3  
332 (Fig. 5E). Indeed, the effective delay between the two inputs to LN3 is 25.3 ms, similar to the delay  
333  $\Delta_{rb} = 23$  ms found for the simple rebound model.

334 The DC tuning of Anurogryllus arises in the last neuron of the full network, in LN4 (Fig. 5E).  
335 LN4 receives excitatory input from the coincidence detector LN3 and feed-forward inhibition from  
336 LN2. The inhibition from LN2 shapes the DC tuning by suppressing responses to song with a DC  
337 of 50% at  $2T_s$  (Fig. 5F): At DCs around 50%, the excitatory input from LN3 is ineffectual because it  
338 overlaps with the strong inhibition from LN2. For higher and lower DCs, inhibition is less potent and  
339 hence the output from coincidence detection prevails and LN4 responds. At lower DCs, inhibition is



**Figure 4: Tuning for pulse rate and duty cycle in the resonate and fire model fitted to Anurogryllus behavior.**

**A** The resonate-and-fire (R&F) model is a spiking neuron model with bidirectionally coupled current (purple) and voltage-like (orange) variables. Inputs currents trigger oscillations with a frequency  $\omega$ . Inputs are excitatory during positive phases and inhibitory during negative phases of the oscillations. If the voltage exceeds a threshold, a spike (grey) is elicited and the current and voltage are reset.

**B** Pulse-pause field (PPF) for the R&F model fitted to Anurogryllus data. Colored lines correspond to the DC and period transects shown in C and D (see legend).

**C** Period tuning of the R&F model for different DCs. Resonant peaks arise at periods at integer multiples of  $T_s$ . The response at  $2T_s$  is attenuated for lower DCs, as in the behavior. Vertical lines correspond to the frequencies shown in D.

**D** DC tuning for three different pulse periods. There is no peak for  $T_s/2$ . At  $T_s$ , the DC tuning is band-pass. At  $2T_s$ , the DC tuning is bimodal, as in the data.

**E** Response traces for the R&F model for songs with different periods (fractions and multiples of  $T_s$ ) and a DC of 33% (stimulus—blue, current—pink, voltage—orange, spikes—grey, see legend). Membrane oscillations and responses are weak at fractions at  $T_s$ . Responses are strong at integer multiples of  $T_s$ . **F** Pulse rate tuning at DC 33%. Shown are the integrals of the stimulus (blue) and spiking response (grey). The current-like (pink) and voltage-like (orange) variables were rectified before integration.

**G, I** Response traces for different DCs at  $T_s$  (G) and  $2T_s$  (I).

**H, J** DC tuning at  $T_s$  (H) and  $2T_s$  (J). Dots mark the stimuli shown in G and I. DC tuning is unimodal at  $T_s$  and bimodal at  $2T_s$ .

Gray boxes in E, G, and I illustrate the stimulus parameters for which traces are shown in the context of the PPF (compare B).

340 weak and offset in time from the excitatory input from LN3. At higher DCs, the excitation from LN3  
341 is stronger and arrives slightly later than the inhibition. In summary, the *Anurogryllus* tuning arises  
342 serially, through two computations in the network model: Rebound and coincidence detection in  
343 LN3 shape the period tuning and feed-forward inhibition from LN2 suppresses responses at wrong  
344 periods and shapes the DC tuning.

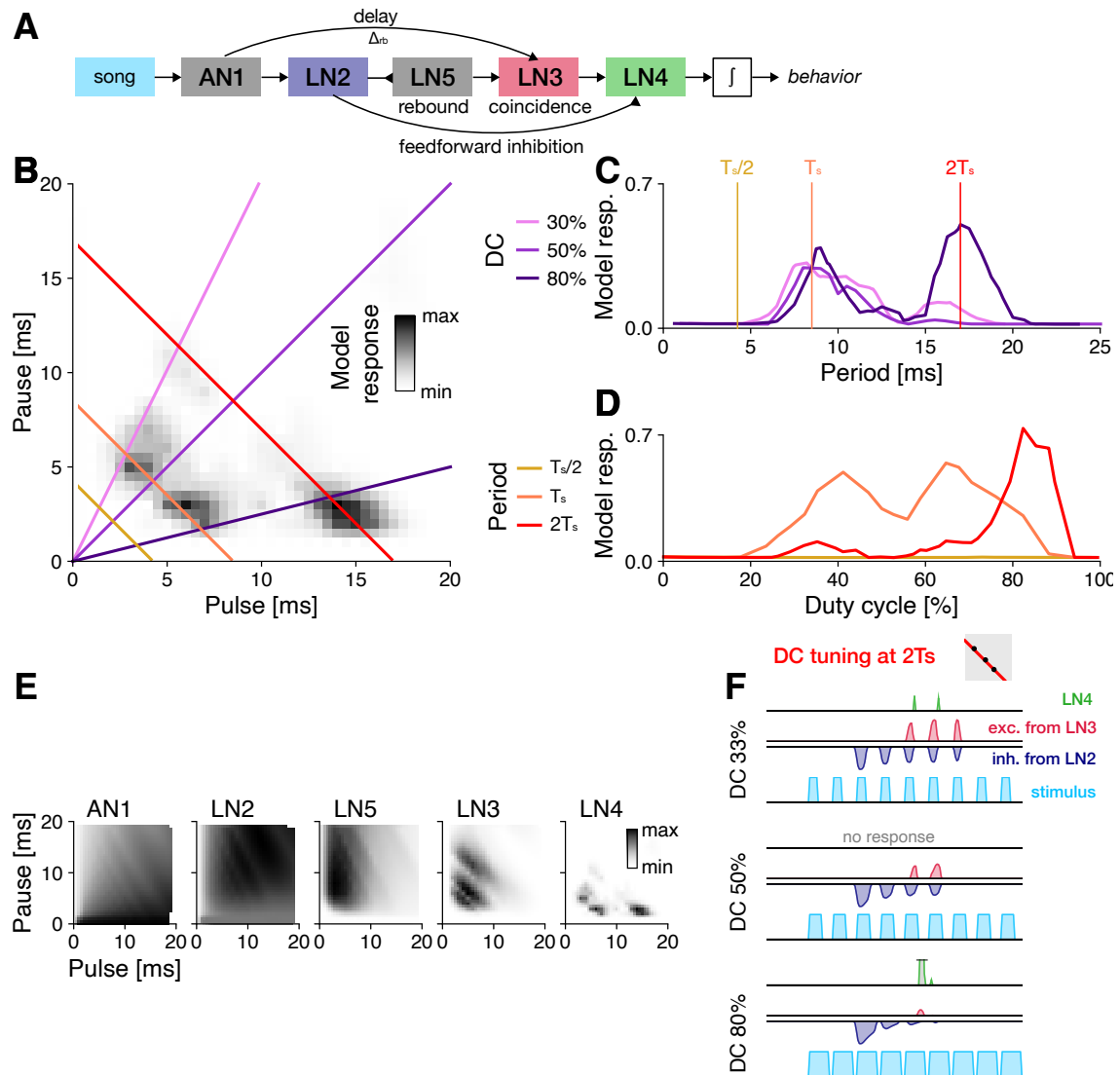
345 To confirm that a mechanism comprising rebounds and feed-forward inhibition is sufficient to  
346 reproduce the *Anurogryllus* behavior, we extended the simple rebound model (Fig. 3) with delayed  
347 feed-forward inhibition (Fig. S2A). We used the parameters of the simple rebound model (Fig.  
348 3) and only fitted the delay and filter properties of the LN2-like input to LN4 (see Methods). This  
349 model is sufficient to reproduce the resonant period tuning (Fig. S2C) and the bimodal DC tuning of  
350 *Anurogryllus* (Fig. S2B–D). The DC tuning arises from the timing of excitatory and inhibitory inputs  
351 to LN4 (Fig. S2E), not from their strengths (Fig. S2F). Responses to a DC of 50% are suppressed  
352 in LN4 because excitation and inhibition arrive at the same time (Fig. S2E). For shorter/longer  
353 DCs, inhibition arrives too early/late to cancel the excitation.

## 354 **Nonlinear computations can accelerate the divergence of song preferences** 355 **through saltatory evolution**

356 Resonant, multi-peaked preference functions as found in *A. muticus* (Fig. 1C–E) may impair  
357 species discrimination, because they produce responses not only to the period of the conspe-  
358 cific song but also to its multiples or fractions. However, resonant recognition mechanisms could  
359 drive the fast co-divergence of song structure and song preference between sister species: Ac-  
360 cording to the standard model of evolution, novel phenotypes evolve through an accumulation of  
361 small genetic changes that induce small phenotypic changes. However, an alternative, saltatory,  
362 model poses that nonlinearities in the mapping from genotype to phenotype can drive sudden  
363 large phenotypic changes (Gould and Eldredge, 1977). Evolutionary developmental biology has  
364 shown that strongly nonlinear developmental programs can give rise to morphological innovations  
365 from small genetic changes (Müller, 2007)—so-called morphological monsters. Resonant song  
366 recognition with responses to disjoint sets of songs is also the result of a highly nonlinear mapping  
367 from network parameters to behavior. If simple mechanisms existed to isolate individual resonant  
368 peaks, then behavioral preferences could jump between these peaks, resulting in sudden large  
369 changes in the female preference that will drive large changes in male song and a rapid isolation  
370 between sister species.

371 Spike-frequency adaptation (SFA) is one mechanism that can isolate individual peaks from a  
372 resonant preference function: SFA is ubiquitous in the nervous system and is also found in the song  
373 recognition network of *G. bimaculatus* (Benda and Hennig, 2008; Schöneich et al., 2015; Clemens  
374 et al., 2021). SFA in combination with the low-pass properties of the neuronal cell membrane  
375 results in a band-pass filter that can be tuned by changing the time constants of the membrane or  
376 of the adaptation current (Benda and Herz, 2003; Benda, 2021). We have implemented a simple  
377 proof-of-principle model to illustrate that SFA can isolate individual peaks from a resonant response  
378 field (Fig. 6B–F). In the example, changes in the membrane time constant of an adapting neuron  
379 can change the relative amplitudes of the individual peaks and thus hide or reveal individual peaks  
380 without creating intermediate ones. This will exert selection pressure on the male song to jump to  
381 the new larger peak of the female preference function. SFA could thus be a mechanism through  
382 which acoustic communication evolves in a saltatory manner: Not by gradual shifting of female  
383 preference and male songs but by jumping of the preferences and songs between relatively fixed  
384 resonant peaks (Fig. 6A).

385 Direct evidence for this hypothesis is difficult to obtain as data on female song preference from  
386 other *Anurogryllus* species does not exist. However, given that female preference and male song  
387 are hypothesized to co-evolve and typically do so in crickets (Kostarakos and Hedwig, 2012; Gabel  
388 et al., 2016; Stout et al., 1983; Doherty and Storz, 1992; Shaw and Herlihy, 2000; Grace and Shaw,  
389 2011), song data could be used as indirect support of our hypothesis. Under the hypothesis, the  
390 pulse periods of males from different *Anurogryllus* species should be close to multiples or fractions  
391 of each other. While song data from the genus *Anurogryllus* is scarce we did find information on the  
392 song from six other *Anurogryllus* species (Table 2), and that data is consistent with our hypothesis  
393 (Fig. 6G). The distribution of song periods of all seven species is trimodal with the modes at  
394 periods of 7.2, 13.4, and 22.8 ms, close to integer multiples of each other. This is consistent with  
395 songs changing in integer steps along resonant peaks. In addition, the first two modes at X and  
396 Y ms are close to the resonant bands of the *A. muticus* preference function (compare Fig. 1D),  
397 with 45% of the song data overlapping with the resonant bands from *A. muticus females*. Under



**Figure 5: A model of the full song recognition network in crickets reproduces the resonant tuning of *Anurogryllus***

**A** Schematic of the full 5-neuron network and internal connections. Pointy and blunt ended arrows indicate excitation and inhibition, respectively. Delay (AN1-LN3), rebound (LN5), and coincidence (LN3) are computations of the core rebound mechanism (Fig. 3). Feed forward inhibition from LN2 to LN4 is crucial for reproducing DC tuning.

**B** The resonant phenotype of *Anurogryllus* recovered with the five neuron model. Colored lines correspond the period and DC transects in D and E.

**C** Traces of the DC transects labeled in B at 33%, 50%, and 80% DC, which each reveal the relative strength of the peaks at  $T_s/2$ ,  $T_s$ , and  $2T_s$ . There is no response at the shortest period ( $T_s/2$ —yellow). At the period of male song ( $T_s$ —orange), DC tuning is band-pass. At the 17 ms period ( $2T_s$ —red), tuning is biphasic, as observed in the behavioral data. Vertical lines correspond to the DCs shown in C.

**D** Traces of the period transects labeled in B, which shows that each peak has unique DC preferences (Compare with the behavioral data in Fig 1 which shows that *Anurogryllus* similarly demonstrates a bandpass preference around the male calling song  $T_s$ , and a preference for high DCs for the  $2T_s$  peak).

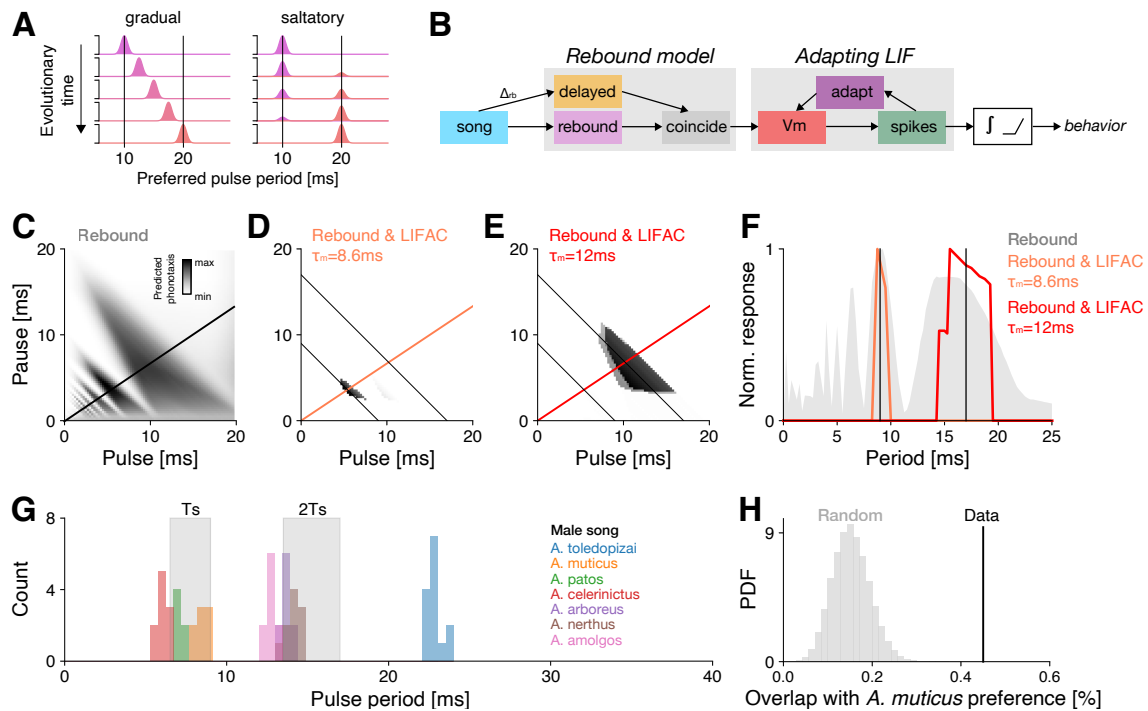
**E** Response profiles of the five neurons in the network.

**F** Example internal network traces for three songs (blue) along the  $2T_s$  period transect at different DC's, showing the interaction of the excitatory coincidence detection output from LN3 (red) and the inhibition from LN2 (blue) to produce the output response in LN4 (green).

Gray boxes in F illustrate the stimulus parameters for which traces are shown in the context of the PPF (compare B).

See also Fig. S2 and 6.

398 a uniform random model, the expected overlap is  $15 \pm 4\%$  and the observed distribution of songs  
 399 is thus unlikely to have arisen by chance ( $p < 10^{-12}$ , Fig. 6H). While this preliminary analysis  
 400 does not provide proof, the results are consistent with the hypothesis that song preference and  
 401 song structure may develop in a saltatory manner by jumping between more or less fixed resonant  
 402 peaks in *Anurogryllus*. In the future, a more comprehensive survey of male song and female  
 403 preference in different *Anurogryllus* species is required to further test the hypothesis.



**Figure 6: Resonances enable saltatory evolution of song preferences**

**A** Evolution of the period preference (top to bottom) in a population under a gradual (left) and saltatory (right) mode. Under a gradual mode, small changes in the preference lead to a shift of the preference over time. Under a saltatory mode, the preference function of individuals jumps to a new peak and that new peak gets fixated without intermediate.

**B** Structure of the rebound model with adaptation. The non-integrated output of the rebound model from Fig. 3 was used to drive a leaky integrate and fire neuron with an adaptation current (LIFAC). The spike output of the LIFAC is then integrated to yield a value proportional to the phonotaxis. A rectifying nonlinearity (relu) is then used to further sharpen the tuning for song.

**C** PPF of the rebound model with resonant peaks used as the input to the LIFAC. The two resonant peaks at  $\approx 9$  ms and  $\approx 17$  ms shown as thin black anti-diagonal lines. The thicker black diagonal line shows the transect at a DC of 66% shown in E.

**D, E** PPFs of the rebound & LIFAC model. The resonant peaks at  $\approx 9$  ms and  $\approx 17$  ms (thin black lines) were isolated by setting membrane time constants  $\tau_m$  to 8.6 (D) and 12.2 ms (E), respectively. The orange and red diagonal lines correspond to the transects at a DC of 66% shown in E.

**F** Period tuning of the models in B–D for a transect through the PPF at a DC of 66%.

**G** Distribution of song periods for seven *Anurogryllus* species. The gray shaded regions depict the responses of *A. muticus* females to the period of the male song ( $T_s$ ) and to twice that period ( $2T_s$ ) (cf. Fig. 1C, D).

**H** Overlap between songs from the 7 species and the resonant bands in the preference function of *A. muticus* females (grey bands in G) in the data (0.45, black line) and under a random uniform model (grey histogram, 100,000 random samples, mean  $\pm$  standard deviation  $15 \pm 5\%$ ). The observed overlap is unlikely to have arisen from that model ( $p < 10^{-12}$ ).



## Discussion

In this paper, we investigated the consequences of the unique song recognition phenotype of *Anurogryllus* for the evolution of acoustic communication among crickets. *Anurogryllus* females respond to three different pulse patterns: pulse patterns matching the period of the male song but also to patterns with twice the period and low or high duty cycle (Fig. 1). Using computational modeling, we tested whether this unusual recognition phenotype in crickets necessitates a corresponding novel recognition mechanism or if the hypothesized shared mechanism observed in other cricket species suffices.

First, to identify elemental computations required for resonant song recognition, we tested simple delay and filter-based network models, alongside a single-neuron model with resonant membrane properties (Figs. 2, 3, 4). While each model could resonate, it was the resonate and fire single-neuron model that best matched the tuning of *Anurogryllus* for both period and DC. That a single-neuron model qualitatively matches the behavior suggests that changes in intracellular properties capable of inducing oscillations of the membrane potential could underlie the evolution of resonant song recognition in *Anurogryllus*.

Critically, we found that a pure rebound mechanism, the core computation of the hypothesized shared mechanism for song recognition in crickets, is insufficient to reproduce the tuning of *Anurogryllus* and that additional computations present in the shared network are necessary (Fig. 5): The core rebound mechanism gives rise only to the resonant period tuning but not the DC tuning, casting doubt on the mother network hypothesis. However, the addition of feed-forward inhibition, present in the full model, recovered the DC tuning profile (Fig. S2). In *G. bimaculatus*, the cricket species in which the network was described, feed-forward inhibition primarily served to refine period tuning (Schöneich et al., 2015), as no resonances appear at the coincidence detection stage of the network in this case. In *Anurogryllus*, it appears to have been coopted to modulate DC tuning by attenuating responses to intermediate DCs. This is in agreement with the fitted mother network model, in which the resonant response arises in two steps: The rebound-based mechanism at the core of the network shapes the period tuning, while feed-forward inhibition shapes the DC tuning. Importantly, in the original network, the feed-forward inhibition only sharpens the period tuning, suggesting that this computation can be re-used in *Anurogryllus* for another function. Overall, our study shows how novel behaviors can arise from the modification of existing intracellular and network computations.

### Mechanisms of resonant song recognition in *Anurogryllus*

In the absence of physiological recordings, computational modeling can be used to constrain hypotheses about the recognition mechanism of *Anurogryllus*. Here, we used two approaches: 1) Minimal models of networks and single-neurons, to identify the computations required to produce the *Anurogryllus* tuning, and 2) a complex network model based on the song recognition network from another species, *G. bimaculatus*, to test about the potential of that network to produce resonant behavior. We identified two mechanisms that can give rise to the resonant song recognition in *Anurogryllus*: A cell-intrinsic mechanism based on oscillatory membrane properties (Fig. 4). And a combination of two network mechanisms: rebound and feed-forward inhibition (Fig. 5, S2).

The resonate and fire neuron, a single-neuron model that was previously used to reproduce resonant song recognition in a katydid (Webb et al., 2007) qualitatively reproduced the pulse rate and DC tuning of *Anurogryllus*. It produced responses to  $T_s$  and  $2T_s$  and exhibited bi-modal DC tuning at  $2T_s$  (Fig. 4). Off-target responses in the model for longer periods could be suppressed by additional computations like a high-pass filter, for instance via adaptation, a computation that is ubiquitous in the mother network (Benda, 2021; Clemens et al., 2020). The resonant membrane properties could arise by changing the expression levels of specific ion channels in any of the neurons of the mother network. For instance, voltage-gated calcium ( $Ca_V$ ) or potassium (KCNQ, HCN) (Ge and Liu, 2016).

We also found that the rebound mechanism in the mother network alone was not sufficient to produce the tuning of *Anurogryllus* (Fig. 3). However, combining the rebound with feed-forward inhibition recreates the period and DC tuning (Fig. S2). In the full network model (Fig. 5A, B), these computations arise in different neurons of the network: First, the rebound produced by LN5 is combined with delayed excitation from AN1 in the coincidence detector LN3 to produce the resonant period tuning. Then, feed-forward inhibition from LN2 shapes the DC tuning in LN4. Crucial for tuning the network are the response delays: from AN1 onto LN3 to tune the preferred periods (Clemens et al., 2021) and from LN2 onto LN4 to tune the DC responses (Fig. S2E, F).

461 Ultimately, determining which of the two proposed mechanisms—single cell or network—generates  
462 the resonant behavior of *Anurogryllus* will require intracellular recordings that detect membrane  
463 oscillations and assess response delays.

## 464 **Resonances are rare because they are undesirable, or because they have** 465 **been missed in experiments**

466 Our analysis of the simple models revealed that resonances can arise easily from common mecha-  
467 nisms like delays or membrane oscillations (Fig. 2, 3, 4). However, multi-peaked response profiles  
468 are known only from two species—*Anurogryllus* and *T. cantans* (Bush and Schul, 2006). This may  
469 reflect selection against resonant tuning, because resonances broaden the female tuning to re-  
470 gions that fall outside of the male calling song, leading to the potential misidentification of mating  
471 partners. While multi-peaked tuning can still enable mate recognition if other signalers do not sing  
472 at the resonant off-target peaks (Amézquita et al., 2011), it is likely that these resonances are sup-  
473 pressed and hidden in many pattern recognition networks, for instance through spike-frequency  
474 adaptation (Fig. 6B–F).

475 However, multi-peaked responses might also be underreported, since their detection requires  
476 a comprehensive and systematic sampling of the stimulus space when quantifying female pref-  
477 erences. Future playback experiments should therefore be designed to ensure the detection of  
478 resonances: Stimuli should not only densely sample different periods but should also do so at  
479 multiple DCs. For instance, a stimulus set that densely samples pulse periods, but at a DC of  
480 50% would have missed the resonant peaks at twice the song period in *Anurogryllus* and *T. can-*  
481 *tans* (Fig. 1G, H). A characterization of the DC tuning also helps differentiate between resonant  
482 mechanisms (Figs. 2, 3, 4): Only the R&F model produces bimodal DC tuning at  $2T_s$ , while  
483 autocorrelation and rebound mechanisms produce unimodal DC tuning. Sweep or chirp stimuli  
484 commonly used in electrophysiology have a changing pulse rate or period are not sufficient for  
485 discriminating models since these stimuli have a constant DC (Narayanan and Johnston, 2007).  
486 Similarly, the presence or absence of responses to odd and even multiples or fractions of the song  
487 period can disambiguate between different mechanisms (Figs. 2, 3, 4): The R&F model responds  
488 only to even multiples of the model's characteristic period, while the simple delay-based models  
489 respond to both even and odd fractions. However, an interpretation of such experiments is com-  
490 plicated by the fact that the behavioral preference is the outcome of multiple computations, in the  
491 case of *Anurogryllus* possibly of a rebound mechanism combined with feed-forward inhibition (Fig.  
492 S2).

## 493 **Resonant song recognition and the evolution of acoustic communication in** 494 **crickets**

495 Overall, our computational approach revealed the capacity of neural networks for change: The  
496 song recognition network described in *G. bimaculatus* consists of a set of elementary computations—  
497 rebounds, coincidence detection, adaptation, feed-forward inhibition—that can give rise to a rich  
498 set of recognition behaviors. This network has the capacity to produce all recognition types known  
499 in crickets: For pulse pause, pulse period, DC, and even multi-peaked resonant tuning of *Anuro-*  
500 *gryllus*. This network could therefore serve as a mother network, that gives rise to the full diversity  
501 of song recognition in crickets. That even a small network, consisting of only 5 neurons can pro-  
502 duce so many diverse behaviors highlights the enormous potential of neural networks to produce  
503 evolutionary novel phenotypes.

504 While simple models of the core rebound, delay, and coincidence detection mechanism only  
505 partially recovered the characteristics of the resonant *Anurogryllus* behavior, insights from the full  
506 model revealed that the function of the pulse recognition network in crickets might include ad-  
507 ditional selectivity for duty cycle via feed-forward inhibition, the inclusion of which enabled the  
508 simple rebound model to replicate the resonant pattern. These results further suggest that the  
509 function of the pulse pattern recognition network in crickets cannot be conceptualized merely as  
510 a rate detector, but that it may additionally select for the duty cycle characteristics of incoming  
511 song, necessitating the inclusion of feed-forward inhibition in even a minimal model for song fea-  
512 ture recognition networks in crickets. More generally, the capacity of neuronal networks to drive  
513 evolutionary change stems in part from the multitude of nonlinear computations at the network and  
514 single-neuron level, which can be coopted to produce new behaviors.

## 515 **Nonlinear computations can drive saltatory behavioral evolution**

516 The resonant mechanism for song recognition in crickets studied here is just one example of the  
517 many nonlinear computations inherent in the neuronal networks that drive behavior. However, they  
518 help illustrate a different view on the evolution of behavior: While the standard model of evolution,  
519 gradualism, assumes phenotypic change through the accumulation of small adaptive changes, an  
520 alternative view poses that large, saltatory change can drive rapid phenotypic change that is then  
521 fixed through selection (Goldschmidt, 1940; Gould and Eldredge, 1977). This saltatory model is  
522 supported by the existence of so-called "morphological monsters", like flies with legs instead of  
523 antennae, which are a symptom of the highly nonlinear genetic networks and programs that drive  
524 morphological development (Müller, 2007). We propose that the highly nonlinear neuronal com-  
525 putations inherent in the brain can also drive saltatory behavioral evolution, and thus behavioral  
526 monsters: Animals with highly unusual behaviors. A saltatory mode of evolution may be most  
527 advantageous if rapid behavioral changes are adaptive, for instance in traits that support species  
528 evolution. The potentially multi-peaked recognition phenotypes driven by resonant mechanisms  
529 can be a trait that allows saltatory changes in song recognition. As one example, we have shown  
530 that spike-frequency adaptation after a resonant recognition network can support this scenario  
531 (Fig. 6B–F) (Benda and Herz, 2003; Benda, 2021). Changes in adaptation parameters can sup-  
532 press one resonant peak and amplify another, driving a switch to a novel preferred pulse period  
533 without intermediates (Fig. 6A). Following the change in female preference will force male songs  
534 to change drastically as well. It is an open question what nonlinear mechanisms might support the  
535 rapid—and possibly saltatory—change of song pattern in the song pattern generators. However,  
536 the same mechanism at work in the female recognition network that gives rise to resonances—  
537 post-inhibitory rebounds, response delays, adaptation, inhibition—can also be found in the song  
538 pattern generators (Schöneich and Hedwig, 2011; Jacob and Hedwig, 2019; Schöneich, 2020).

539 While the idea of saltatory evolution is theoretically intriguing, it also generates testable hy-  
540 potheses on the statistics of male song pattern and female song preference within a species group.  
541 Assuming that male's pulse pattern and the female preference preference co-evolve (Kostarakos  
542 and Hedwig, 2012; Gabel et al., 2016; Stout et al., 1983; Doherty and Storz, 1992; Shaw and  
543 Herlihy, 2000), saltatory evolution by jumping between fixed resonant peaks would lead to multi-  
544 modal distributions of song parameters and song preferences in closely related species. While by  
545 no means ultimate since it is based on a small set of song data, our preliminary finding that the  
546 songs of seven species in the genus *Anurogryllus* are trimodally distributed is consistent with this  
547 prediction. To further test our hypothesis, songs from more species under consideration of phy-  
548 logenetic data are required. As is female preference data, which would show that females from  
549 different species also show either multi-peaked responses like *A. muticus* (Fig. 1C, D) or fall on  
550 a small number of peaks. In addition, if a species is "caught in the act" of transitioning from one  
551 peak to another, we would expect bimodal distributions of female preferences and/or male song  
552 parameters.

553 Given the existence of nonlinear computations in neural networks, the potential for saltatory be-  
554 havioral evolution exists in every system and it might drive evolution whenever sudden phenotypic  
555 changes are adaptive.

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## 561 **Contributions**

- 562 • Conceptualization - WM, MH, JC
- 563 • Animals and behavioral experiments - BE, MH
- 564 • Modeling and analysis - WM, JC
- 565 • First draft - WM, JC
- 566 • Feedback on draft - BE, MH

## 567 Methods

### 568 Animals

569 Behavioral experiments were performed with *Anurogryllus muticus* from the same colony as used  
570 in (Erregger et al., 2018). The progeny of individuals caught on Barro Colorado Island in Panama  
571 were reared to adulthood at the Department of Zoology at the University of Graz in Austria and  
572 held at 25–28 °C with *ad libitum* food and water. Starting with the second or third instar, individuals  
573 were separated from the colony and placed in individual plastic boxes.

### 574 Male song recording and analysis

575 Individuals were placed in an array of separate boxes (mean temperature  $24.9 \pm 1.0^\circ\text{C}$  SD) for  
576 a duration of 16–24 hours. Each box was equipped with a microphone and isolating foam to en-  
577 sure acoustic isolation. Using customized software (LabVIEW 7, National Instruments, Austin, TX,  
578 USA), the microphone (TCM 141 Conrad; Conrad Electronic, Germany) in each box was scanned  
579 for 800 ms at a time with a sampling rate of 100 kHz and a male was recorded for 20 s if it produced  
580 sound during that 800-ms interval (Hennig et al., 2016). The song carrier frequency was deter-  
581 mined from the spectral peak of the raw waveform signal. For analysis of the temporal pattern,  
582 the normalized envelope of the song signal was computed after signal rectification by squaring  
583 and low-pass filtering at 200 Hz (equivalent to a temporal resolution of 2.5 ms). Temporal param-  
584 eters such as pulse and pause duration were calculated when the envelope crossed or fell below  
585 a threshold value at 10–15% of the signal envelope.

586 As a preliminary test of our hypothesis that resonance might drive saltatory evolution, we ex-  
587 amined the pulse periods of calling songs from six more species of the genus *Anurogryllus* from  
588 the literature (Table 2). Aside from *A. muticus*, only *A. toledopizai* and *A. patos* included individual  
589 specimen measurements. For each of the species for which only one pulse period was re-  
590 ported (*A. celerinictus*, *arboreus*, *nerthus*, *amolgos*), distributions of 10 songs were generated by  
591 sampling from a Gaussian with the species' pulse period as the mean and the standard deviation  
592 from *A. muticus*.

593 Together with the species with individual specimen measurements, we find that roughly 45% of  
594 these 68 songs fall within either the  $T_s$  or  $2T_s$  period harmonics of *A. muticus* ( $6.5 \leq T_s \leq 9$  ms or  
595  $13.5 \leq 2T_s \leq 17$  ms). To test whether this distribution of songs deviates from a random uniform  
596 distribution we simulated 100,000 trials in which 68 random songs drawn were from a uniform  
597 distribution between 0 and 40 ms and calculated for each of the 100,000 random distributions  
598 their overlap with the  $T_s$  or  $2T_s$  harmonic bands from *A. muticus*. The resulting distribution of  
599 100,000 probabilities was then compared to the observed overlap of 45% and the probability of  
600 obtaining an overlap of 45% or greater was estimated by approximating the random distribution as  
601 a Gaussian to be  $p < 10^{-12}$ .

Species	Pulse period (mean $\pm$ std)	Sample Size (N)	Source
<i>A. muticus</i>	8.5 $\pm$ 0.3 ms	8	Erregger et al., 2017
<i>A. toledopizai</i>	22.8 $\pm$ 0.47 ms	14	Redü and Zefa, 2017
<i>A. patos</i>	7.2 $\pm$ 0.22 ms	6	Redü and Zefa, 2017
<i>A. celerinictus</i>	5.8 $\pm$ * ms	23	Walker, 1973
<i>A. arboreus</i>	13.5 $\pm$ * ms	19	Walker, 1973
<i>A. nerthus</i>	14.08 $\pm$ * ms	*	Walker, 2015
<i>A. amolgos</i>	12.65 $\pm$ * ms	*	Walker, 2015

**Table 2: Song periods and their distribution from seven *Anurogryllus* species.** \* indicates that this statistic was not reported in the source literature.

### 602 Female preference functions

603 Female preference was tested using a trackball system as described in (Hennig et al., 2016). Fe-  
604 males, mounted to a metal rod, were placed on a hollow Styrofoam sphere (diameter: 100 mm,  
605 weight: 1.2–1.8 g) supported by an air stream between two perpendicularly placed loudspeakers  
606 (Piezo Horntweeter PH8; Conrad Electronic) in a wooden box with sound absorbing foam. Each

607 loudspeaker was calibrated with a Bruel and Kjaer 2231 sound level meter and a half-inch con-  
608 denser microphone (Bruel and Kjaer 4133 relative to 0.02 mPa, fast reading) at the top of the  
609 sphere where the female cricket was placed during experiments.

610 Digitally stored sound signals were transmitted from a hard disk by a D/A-board (update rate:  
611 100 kHz, PCI 6221; National Instruments, Austin, TX, USA) to a digitally controlled attenuator  
612 (PA5; Tucker-Davis, Alachua, FL, USA), amplified (Raveland; Conrad Electronic) and broadcasted  
613 through the speakers. The longitudinal and lateral movements of the sphere were recorded by  
614 either a single optical sensor (Agilent ADNS-2051; Agilent Technologies, Santa Clara, CA, USA)  
615 at the bottom of the half-sphere or by two sensors (ADNS-5050; Avago Technologies, San Jose,  
616 CA, USA) with a focusing lens positioned laterally at an angle of 90°.

617 A silent control was used to monitor baseline walking activity, and a continuous tone was used  
618 to control for motivation and selectivity of female responses. At the beginning and the end of  
619 each test session, a species-specific, attractive song signal was presented to control for possible  
620 changes in phonotactic motivation during a session. For each test signal, the lateral deviation of a  
621 female during signal presentation for each of the two speakers was averaged and normalized with  
622 respect to the attractive control signal. The resulting phonotactic scores were therefore typically  
623 between 0 (no orientation towards the sound signal) and 1 (strong orientation towards the signal),  
624 although negative scores (orientation away from the signal) and scores higher than 1 (orientation  
625 towards signal stronger compared to control stimulus) were possible. Test signals and controls  
626 were presented at 80 dB sound pressure level. All tests included the four control stimuli (silent,  
627 continuous tone, and an attractive stimulus at the beginning and end of a test) and eight test stimuli  
628 (total duration was 29 min per test), and were performed at 24°C.

629 Phonotaxis values were measured for 75 artificial pulse trains, split into 10 playlists. Each  
630 playlist was tested with 3-8 females and the phonotaxis values for each stimulus were averaged  
631 over the females (Fig. S1). All stimulus parameters, phonotaxis values, and number of animals  
632 are listed in a supplemental data file. From the 75 average phonotaxis values, we generated a  
633 two-dimensional preference function using natural neighbor interpolation implemented in `metpy`  
634 (URL: <https://github.com/Unidata/MetPy>). The preference function covered pulse and pause  
635 durations between 0 and 20 ms, with a resolution of 0.1 ms. Negative phonotaxis values in this  
636 interpolated preference functions were set to 0.

## 637 Modeling

### 638 Stimulus and response data

639 Song pulses were constructed as rectangular boxes with an amplitude of 1. While natural pulse  
640 trains in *Anurogryllus* last for many seconds, the models tested here have dynamics on the timescale  
641 of a few tens of milliseconds. To speed up simulations, we therefore used pulse trains with a du-  
642 ration of 400 ms and omitted onset and offset transients when translating the model output to  
643 predicted phonotaxis (see below). The stimulus set contained pulse trains with all combinations  
644 of pulse and pause durations between 0–20 ms sampled on a grid with an interval of 0.5 ms, to-  
645 tallying  $(20/0.5)^2 = 1600$  stimuli. As the fitting target, we used the two-dimensional preference  
646 function from *Anurogryllus* females obtained by interpolating the experimental phonotaxis values  
647 as described above, but on a grid with a step size of 0.5 ms.

### 648 Predicting phonotaxis score from model responses

649 The predicted phonotaxis score,  $p$ , is given by the average model response  $r(t)$  over the stim-  
650 ulus duration  $D_s$ , excluding the first 25 ms and the last 10 ms to reduce the impact of response  
651 transients:  $p = 1/(D_s - 35ms) \int_{25ms}^{D_s-10ms} r(t) dt$ .

### 652 Model fitting

653 The models were fitted using the Nelder-Mead method implemented in `scipy.optimize.minimize`,  
654 by minimizing the mean-squared error between the interpolated phonotaxis values from the data  
655 and the model response. If not stated otherwise, initial values for all parameters were set using a  
656 vector of initial conditions chosen manually to speed up fitting. Fits were run multiple times from  
657 slightly different initial conditions to avoid getting stuck in local minima. The presented parameter  
658 values are from models with the lowest error. The model parameters for the simple models are  
659 listed in Table 3 and for the full network in Table 4. The code and parameters for running all models  
660 can be found at <https://github.com/janclemenslab/anurogryllus-resonance>.



## 661 Autocorrelation model

662 In the autocorrelation model (Fig. 2A), the stimulus  $s(t)$  is delayed by  $\Delta_{ac}$ ,  $s_{\Delta}(t) = s(t - \Delta_{ac})$ .  
663 A coincidence detector then multiplies  $s(t)$  and  $s_{\Delta}(t)$  and scales the result with a gain factor  $g_{ac}$ :  
664  $r(t) = g_{ac} \cdot s(t) \cdot s_{\Delta}(t)$ . We did not add a nonlinearity to the output  $r(t)$ , like a sigmoidal, prior  
665 or after integration, since it did not produce quantitatively different predictions during fitting. The  
666 autocorrelation model was simulated with a resolution of 10 kHz.

## 667 Rebound model

668 The rebound model extends the autocorrelation model by inverting and filtering one of the two  
669 paths the stimulus takes before coincidence detection to produce offset responses at the end  
670 of each pulse:  $s_F(t) = \int_0^T -s(t - \tau) \cdot h(\tau) d\tau$ . The filter  $h(\tau)$  consists of two lobes, defined as  
671 rectangular windows: An inhibitory lobe with negative gain  $g_i$  and duration  $T_i$ , followed by an  
672 excitatory lobe with positive gain  $g_e$  and duration  $T_e$ . The positive response components in  $s_F(t)$   
673 corresponding to the rebound are isolated using a rectifying linear function:  $s_R(t) = f(s_F(t))$ , where  
674  $f(x) = 0$  if  $x \leq 0$ , and  $f(x) = x$  if  $x = 0$ . The coincidence detector then multiplies  $s_R(t)$  and  $s_{\Delta}(t)$ :  
675  $r(t) = s_R(t) \cdot s_{\Delta}(t)$ . The rebound model was simulated with a resolution of 4 kHz to accelerate the  
676 fitting process.

## 677 Rebound model with feed-forward inhibition

678 The rebound model with feed-forward inhibition extends the simple rebound model by including an  
679 additional inhibitory connection to the basic rebound model following coincidence detection (See  
680 Fig S2A). The added inhibitory path from stimulus to output (LN4) contains a bi-phasic filter with  
681 rectangular negative and positive lobes (similar to the filter in the rebound model) and a delay. The  
682 negative components of the output of the bi-phasic filter were then used as an inhibitory input to an  
683 LN4-like output neuron. The LN4-like neuron combines the inputs from the coincidence detector  
684 and the feed-forward inhibitory paths. To obtain the predicted phonotaxis value for a given stimulus,  
685 the output of the LN4-like neuron was passed through a rectifying linear function with threshold  
686  $\theta_{relu} = 0$  and a linear gain  $g_{relu} = 1$  and then integrated. When fitting this model, the parameters  
687 of the simple rebound model fitted previously were kept fixed and only the additional parameters  
688 for the feed-forward inhibition branch (the delay time and the gain and duration of the inhibitory  
689 and excitatory lobe) were adjusted.

## 690 Resonate-and-fire neuron

691 The resonate-and-fire model was implemented following Izhikevich (2001):

$$\frac{dx}{dt} = b * x - \omega * y + g_s * s(t)$$

692

$$\frac{dy}{dt} = \omega * x + b * y$$

where  $x$  is a current-like variable,  $y$  is a voltage-like variable,  $b$  is the damping factor,  $\omega$  is the  
intrinsic frequency,  $s(t)$  is the song input and  $g_s$  is the gain of the song input. If  $y$  exceeds the  
threshold  $y_{threshold} = 1$ , a spike with amplitude  $g_{rb}$  is elicited and current and voltage are reset to  
 $x_{reset} = 0$  and  $y_{reset} = 1$ :

$$y = \begin{cases} x = x_{reset} \text{ and } y = y_{reset}, & \text{if } y \geq y_{threshold} \\ y, & \text{otherwise} \end{cases} \quad (1)$$

693 The differential equations were numerically integrated using the Euler method with a time step  
694 of 0.1 ms.

## 695 Full model of the song recognition network in *G. bimaculatus*

696 To test whether the song recognition network from *G. bimaculatus* described in Schöneich et al.  
697 (2015) can reproduce the resonant behavior of *Anurogryllus*, we used the model of the network  
698 from Clemens et al. (2020). This model was fitted to reproduce the response dynamics and the  
699 tuning of all neurons in the network using electrophysiological recordings from *G. bimaculatus* for a



Model	Parameter name	Parameter value
Autocorrelation	delay $\Delta_{ac}$	17.0 ms
	output gain $g_{ac}$	0.21
Rebound	delay $\Delta_{rb}$	22.93 ms
	filter inhibitory gain $g_i$	0.045
	filter inhibitory duration $T_i$	5.06 ms
	filter excitatory gain $g_e$	0.1
	filter excitatory duration $T_e$	2.00 ms
Rebound with feed-forward inhibition (remaining parameters were taken from the rebound model)	delay $\Delta_{ffi}$	7.29 ms
	filter inhibitory gain $g_{fi}$	1.01
	filter inhibitory duration $T_{fi}$	2.43 ms
	filter excitatory gain $g_{fe}$	0.63
	filter excitatory duration $T_{fe}$	2.45 ms
Resonate and fire	frequency $f_{r\&f} = \omega/2/\pi$	109.34 Hz
	damping $b$	-0.0005
	input gain $g_s$	0.027
	output gain $g_{rb}$	0.0025

**Table 3: Parameters of the simple models fitted to reproduce the Anurogryllus preference function.**

700 large set of pulse train stimuli (Kostarakos and Hedwig, 2012; Schöneich et al., 2015). The forty-  
701 five parameters in the network model were fitted using the Nelder-Mead optimization algorithm,  
702 by minimizing the mean-square error between experimental and predicted phototaxis values (see  
703 Table 4 for the fitted parameters) using the parameter values found for *G. bimaculatus* as a start  
704 point. Several rounds of optimization were required to converge on the given parameter set, with  
705 Gaussian-distributed noise added to all parameters at the start of the initial optimization rounds to  
706 avoid undesirable local minima. Model fitting often yielded models that reproduced the tuning of  
707 Anurogryllus with only transient responses at the onset of the pulse train. Given that Anurogryllus  
708 song lasts multiple seconds and elicits phonotaxis throughout, we deemed these solutions physio-  
709 logically unrealistic. We therefore added the constraint that responses of AN1 in the model should  
710 spike throughout the stimulus for pulse trains with conspecific parameters.

Cell	Component	Parameters
AN1	Filter excitatory lobe	(Gaussian) width $\sigma=3.88$ , duration = 7.59 ms, input delay = 2.26 ms
	Filter inhibitory lobe	(Gaussian) width $\sigma = 3.81$ , gain $\gamma = 0.87$ , duration 293.04 ms
	Nonlinearity	(Sigmoidal) slope = 10.33, shift = 0.62, gain = 1.19, baseline = -0.29
	Adaptation	(Divisive normalization) timescale $\tau = 9999.93$ ms, strength $w = 85.75$ , offset $x_0 = 1$
LN2	Input from AN1M	Delay = 7.59 ms, gain = 1.93
	Filter excitatory lobe	(Gaussian) width $\sigma = 9.76$ , duration = 11.87 ms, gain = 0.59
	Filter inhibitory lobe Nonlinearity	(Exponential) decay $\tau = 15.87$ ms, duration N = 1000 ms (Rectifying) threshold = 0, gain = 4.22
LN5	Input from LN2	Delay = 13.13 ms, gain = .43
	Postsynaptic filter	(Differentiated Gaussian) width duration N = 8.94 ms, gain of the excitatory lobe = .41
	Postsynaptic nonlinearity	(Rectifying) threshold = 0, gain = 0.57
	Rebound filter excitatory lobe	(Gaussian) width $\tau = 0.02$ , duration = 5.18 ms, gain = -0.007
	Rebound filter inhibitory lobe	(Exponential) decay $\tau = 17.29$ ms, gain = 6.5 duration N = 1000 ms

	Nonlinearity	(Rectifying) threshold = 0, gain = 0.006
LN3	Input from AN1 Input from LN5	Delay = 16.59 ms, gain = 0.65 Delay = 9.67 ms, gain = 43.73
	Postsynaptic nonlinearity Adaptation	(Rectifying) threshold = 0.24, gain = 6.82 (Divisive normalization) timescale $\tau = 1463.98$ ms, strength $w = .16$
	Nonlinearity	(Rectifying) threshold = 5.1, gain = 3.51
LN4	Input from LN2 Input from LN3 Nonlinearity	Delay = 11.44 ms, gain = -58.26 Delay = 7.15 ms, gain = 3.75 (Rectifying) threshold = -0.003, gain = 6.82

**Table 4: Parameters of the 5 neuron "mother network" model fitted to reproduce the Anurogryllus preference function.**

## 711 Modeling jumps between resonant peaks with spike-frequency adaptation

712 To demonstrate that individual resonant peaks can be isolated from a resonant response field, we  
713 added to the rebound model fitted to the Anurogryllus data (Fig. 3, same parameters as in Table  
714 3) a leaky integrate and fire neuron with an adaptation current (LIFAC) using the code published  
715 with Benda (2021). The LIFAC model is driven by the non-integrated output of the rebound model  
716 and acts as band-pass filter, because it combines the low-pass properties of a cell membrane and  
717 high-pass properties from adaptation (Benda and Herz, 2003). The total spike output from the  
718 LIFAC model for each stimulus is passed through a rectifying linear function with threshold  $\theta_{relu}$   
719 and a linear gain  $g_{relu} = 1$ , to compute the predicted phonotaxis value.

720 The LIFAC neuron responds to a current input  $I$  by increasing the membrane potential  $V$  from  
721 which an adaptation current  $A$  is subtracted:

$$\tau_m \frac{dV}{dt} = -V + I - A \quad (2)$$

$$\tau_{ada} \frac{dA}{dt} = -A \quad (3)$$

722 with time constants of the membrane and of adaptation,  $\tau_m$  and  $\tau_{ada}$ , respectively. If the voltage  
723  $V$  reaches the spiking threshold  $V_{thres}$ , a spike is elicited, and  $V$  is reset to  $V_{reset}$  and the adaptation  
724 current strength  $A$  is incremented by  $\alpha$ :

$$V = \begin{cases} V_{reset} \text{ and } A = A + \alpha, & \text{if } V \geq V_{thres} \\ V, & \text{otherwise} \end{cases} \quad (4)$$

725 Each spike initiates a refractory period  $\tau_{ref}$ , during which both  $V$  and  $A$  are fixed to their reset  
726 values.

Period	Parameter name	Parameter value
shared	Spike Refractory Period $\tau_{ref}$	1 ms
	Adaptation Time Constant $\tau_{ada}$	5 ms
	Adaptation Strength $\alpha$	10 mV
	Spike Threshold $V_{thres}$	0.5 mV
4 ms	Membrane Time Constant $\tau_m$	4 ms
	Threshold $\theta_{relu}$	125 spikes
8 ms	Membrane Time Constant $\tau_m$	8.8 ms
	Threshold $\theta_{relu}$	72 spikes
16 ms	Membrane Time Constant $\tau_m$	12 ms
	Threshold $\theta_{relu}$	0 spikes

**Table 5: Parameters of the rebound model with adaptation shown in Fig. 6G.**

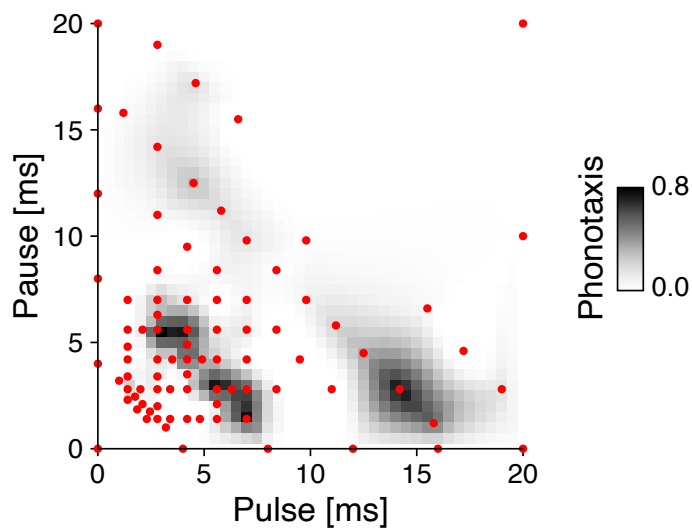
The first four parameters in the table are shared between all variants of the model. Resonant peaks at 4, 8, and 16 ms are isolated by adjusting the membrane time constant,  $\tau_m$ , and the threshold of the rectifying linear function,  $\theta_{relu}$ .

## Supplemental Information

Peak	Females tested	p-value
$T_s/2$ (4.5 ms)	7	0.058
$T_s$ (8.5 ms)	7	0.006
$2T_s$ (17 ms), high DC	4	0.002
$2T_s$ (17 ms), low DC	7	0.007

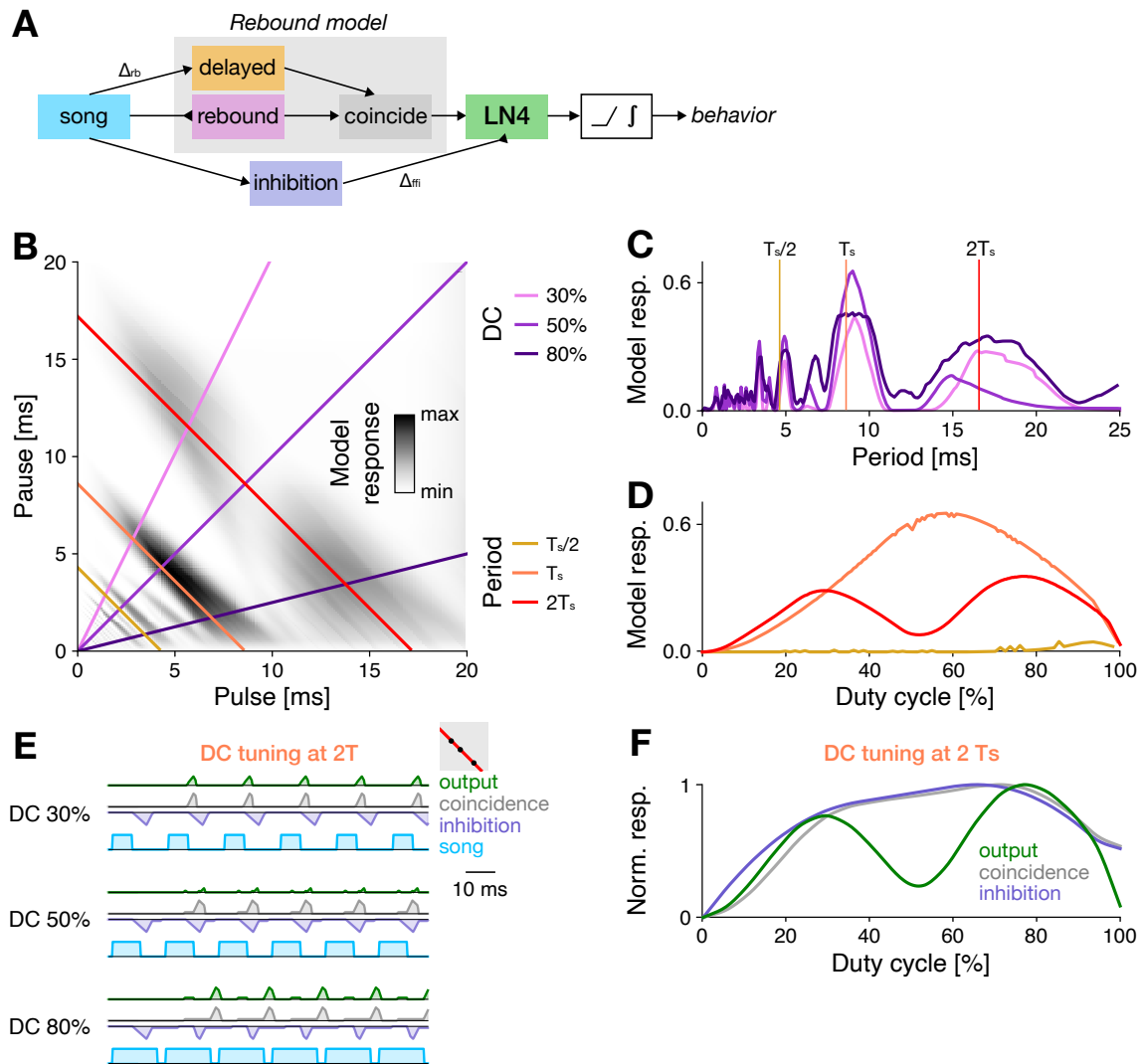
**Table S6: Statistical tests for each peak in the *Anurogryllus* phenotype (Related to Fig. 1).**

P-values were obtained from a paired one-sided t-test testing the hypothesis that the responses of the individuals to songs at the peak are greater than a silent control. All peaks, except for the peak at  $T_s/2$ , are significant. The broad peak at  $2T_s$  for low DC was evaluated using two points within this peak for which different sets of females were tested. The stimuli for these low DC points have either a pause of 12.5 ms and a duration of 4.5 ms, or a pause of 11.2 ms and a duration of 5.8 ms. The high DC condition for  $2T_s$  was evaluated at a pause of 2.8 ms and a duration of 14.2 ms.



**Figure S1: Pulse train stimuli used for estimating the pulse-pause field (PPF) (Related to Fig. 1).**

Individual pulse trains for which phonotaxis values were measured are shown as red dots. The PPF (color coded, see color bar) was obtained by natural neighbor interpolation of the phonotaxis values on a dense 41x41 grid (all combinations of pulses and pauses between 0 and 20 ms with a step size of 0.1 ms). Phonotaxis values at the boundaries (pulse or pause 0 ms) were set to 0.



**Figure S2: A combination of rebound and feed-forward inhibition are sufficient to create the tuning of Anurogryllus. (Related to Fig. 5).**

**A** Schematic of the rebound model with delayed feed-forward inhibition. An LN4-like neuron receives input from the coincidence detector of a rebound model and from inhibition. The output of

**B** PPF illustrating the responses produced by the modified rebound model fitted to behavioral data from Anurogryllus (Fig. 1C, see color bar), demonstrating the restored bimodal shape of the 17 ms period transect. Colored lines correspond to the DC and period transects shown in C and D.

**C** Period tuning of the model for different DCs.

**D** DC tuning for three different pulse periods, corresponding to  $T_s/2$ ,  $T_s$ , and  $2T_s$ . The curves indicate bandpass preference around the male calling song  $T_s$ , and bimodal DC tuning for the  $2T_s$  peak. Vertical lines correspond to the DCs shown in C.

**E** Example traces showing how the delay timing of inhibition (blue) interacts with the coincidence detection output (grey) to produce bimodal tuning along the  $2T_s$  17 ms period transect. Inhibition at 50% DC coincides with the timing of the coincidence detection output, fully suppressing responses.

**F** DC tuning of the rebound output (grey) vs the feed-forward inhibition (blue) for the  $2T$  17 ms transect, which produces the final bimodal tuning (green) as observed in the behavior data.

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